

Anticancer Effect of Bovine Lactoferrin on Breast Cancer Cell Line MCF7 and the Evaluation of *Bax* and *Bak* Genes Expression Involved in Apoptosis



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ABSTRACT

Background: Lactoferrin is a glycoprotein with antimicrobial, antioxidant, immune-modulating, antiviral, and most importantly anticancer properties. In the present study, the effect of lactoferrin on breast cancer cell growth and the expression of *Bax* and *Bak* genes are evaluated.

Materials and Methods: MCF7 cells were cultured in a 96-well plate with 1×10^5 cells in each well. Different lactoferrin concentrations of 0, 50, 300, 600, and 800 $\mu\text{g/mL}$ were added to each well in three replicates and the well was incubated for 24 hours. After treatment, cell survival was measured using the MTT assay. To determine the level of expression of *Bax* and *Bak* genes, the cells were treated with lactoferrin concentrations of 0, 50, and 800 $\mu\text{g/mL}$ in 2 replicates for 24 hours. Then RNA extraction was performed and cDNA was synthesized immediately and the expression of the genes in the presence of beta-actin reference gene and cyber-green fluorescence color was investigated with real-time reactions.

Results: The cells viability in lactoferrin concentrations of 0, 50, 300, 500 and 800 $\mu\text{g/mL}$ were 100%, 94%, 83%, 62%, and 32%, respectively. The expression level of the *Bax* gene at a concentration of 50 μg increased by 2.71 times and in 800 μg concentration decreased by 0.88 times. Also, the expression level of the *Bak* gene at concentrations of 50 and 800 μg increased by 1.23 and 1.0 fold, respectively. Statistical analysis of the data indicated that the expression levels of two genes at a concentration of 50 $\mu\text{g/mL}$ of lactoferrin significantly increased ($P < 0.01$), compared to the control. The significance level in this study was set at $P < 0.05$.

Conclusion: In this study, lactoferrin showed a growth inhibitory effect on breast cancer cells and increased the expression of *Bax* and *Bak* genes involved in apoptosis at a concentration of 50 $\mu\text{g/mL}$.

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Introduction

Cancer refers to the uncontrolled growth and abnormal development of cells in the body. The most common cause of death due to cancer in women is breast cancer [1]. Despite advances in cancer treatment, there is a sense of the need for anti-cancer agents that increase cellular resistance.

Lactoferrin is a multi-function protein that performs several biological functions and has antibacterial, antiviral, antifungal, anti-inflammatory, anti-tumor, antioxidant, and immunological activities [2]. Antitumor activity of lactoferrin in cancer cells has been shown in various tissues, such as pharynx, lung, bladder, liver, colon, stomach, and breast [3-6]. It has also been reported that lactoferrin induces apoptosis in the cancers of the colon, lung, stomach, esophagus, and breast [7-10].

Lactoferrin can induce apoptosis through intracellular and extracellular pathways in cancer cells [11]. The extracellular mechanism is related to its interaction with cell membrane receptors [12]. And the intracellular mechanism is associated with cell apoptosis and cell cycle arrest [13]. The mechanism of lactoferrin cytotoxic actions like membrane disruption, apoptosis, and cell cycle arrest have been reported in different cancer cells [14-16].

One of the important functions of lactoferrin is the regulation and strengthening of the immune system. The immune system cannot destroy cancer cells since tumor cells have a suppressive effect on the immune system. Lactoferrin can activate the immune system in two ways; on the one hand, it supports the proliferation, differentiation, and activation of the immune system cells and thus boosting the immune system and on the other hand, it inhibits the inflammatory factors [17]. Lactoferrin increases the amount of anti-inflammatory interleukin-10 and cytokine-mediated stimulation of natural killer (NK) cells that play an important part in the defense against tumor growth while cancer cells inhibit them [17, 18]. Lactoferrin affects NK cells activation that leads to cancer cell lysis [11].

Bax and *Bak* genes are pro-apoptotic members of the Bcl-2 family needed to program cell death. Dimerization of *Bax* and *Bak* are crucial for the activation of mitochondria resulting in a change in the mitochondrial membrane potential, production of reactive oxygen species, and release of cytochrome C, which activates a downstream caspase program [19-21].

The inducible expression following a death stimulus translocation of Bax into the mitochondria where it becomes an integral membrane protein and is cross-linked as a dimer. Bak is in the outer membrane of the mitochondria that inhibitors of apoptosis prevent it from oligomerization. When this inhibition is removed Bak like Bax forms a dimer and promotes membrane permeability [22, 23]. According to previous studies, the up-regulation of Bax and Bid via lactoferrin could induce apoptosis in stomach cancer cell line SGC-7901 [10]. Lactoferrin induces apoptosis in Jurkat T leukemia cells and MDA-MB-435 breast carcinoma cells by the induction of caspases 2, 3, and 9 activities [24]. Bak can induce apoptosis in various mammalian cells and a decrease in Bak protein expression was reported in primary colorectal adenocarcinomas and gastric adenocarcinomas when compared with the normal tissue [25, 26]. Since Bak and Bax can form a heterodimer with anti-apoptotic Bcl-2 and Bcl-xL and induce apoptosis, Bak maybe a tumor suppressor in some cancers [27, 28]. Therefore, reagents that affect the expression of Bak or Bax can control cancer cell growth.

In this study, the effect of various concentrations of lactoferrin on the growth of breast cancer cell lines (MCF-7) and expression of two apoptotic genes, *Bax* and *Bak*, were investigated.

Materials and Methods

Determination of lactoferrin concentration and purity

Lactoferrin was extracted from cow milk colostrum and purified using cation exchange chromatography [7]. The purity of lactoferrin was checked with 10% SDS-PAGE (sodium dodecyl sulfate-polyacrylamide gel electrophoresis). The concentration of purified protein was determined by the Bradford method.

Cell culture and treatment with lactoferrin

MCF-7 human breast cancer cell line was purchased from the Pasteur Institute (Iran). MCF-7 cells were cultured in RPMI-1640, containing 20% FBS, and incubated at 37°C with 5% CO₂.

After the growth of MCF-7 cells and reaching 70% confluency, the cells were transferred to a 96 -well plate with 5×10³ cells in each well and incubated for 24 hours. Then, the different concentrations of lactoferrin (0, 50, 300, 600, and 800 µg/µL) were treated in three replicates.

MTT assay

To measure the effect of lactoferrin on the growth inhibition of the cancer cells, MTT assay was performed, according to Sigma protocol, and the absorbance was measured at 490 nm using ELISA reader (Stat Fax 2100, Belgium). Primers were designed with Gene Runner and synthesized by Gene-Fanavar. Nucleotide sequences used for the design of probe-primers were retrieved from the NCBI database and the designed probe-primers were aligned by BLAST to confirm gene specificity.

RNA extraction

The grown cells on the 6-well plate were treated with three different concentrations of lactoferrin containing 0, 50, and 800 µg/mL for 24 hours. Then, the total RNA was extracted using the EZ-10 spin column total RNA mini-prep kit (Takara, Japan) according to the manufacturer's manual. The quality of extracted RNA was checked up on agarose gel electrophoresis.

The complementary DNA (cDNA) was synthesized using the Revert Aid first-strand cDNA synthesis kit (Takara, Japan). The quality and quantity of cDNA were analyzed by agarose gel electrophoresis and spectrophotometer, respectively. The cDNA was stored at -20°C.

Real-time PCR

At first, the PCR reaction was done for the evaluation of the amplification of *Bax* and *Bak* genes using specific primers. The specific primers were designed based on the BH3 conserved region of the two genes and β -actin considered as a reference gene for data normalization (Table 1). The LightCycler FastStart DNA Master PLUS SYBR Green I kit (Takara, Japan) was used for a real-time PCR reaction. The reaction was prepared in final volume 25 µL contained, cDNA as template, specific primers, TB green, ROXI, and ddH₂O. The thermal cycling of real-time PCR was the first denaturation at 95°C for 10 min; followed by 40 cycles, including denaturation at 95°C for 30 seconds; annealing at 59°C for 30 s; extension at 72°C for 45 s, and the final extension at 72°C for 10 min. The reaction was performed in the Step-One Plus™ real-time PCR (Australia).

All samples containing different concentrations of lactoferrin treatment and reference genes were amplified in triplicate. After amplification of the genes, data analysis was performed using $2^{-\Delta\Delta CT}$ methods as follows: ΔCT (Bak or Bax genes) = Ct (Bak or Bax genes) - Ct (β -actin), $\Delta\Delta CT$ = ΔCT (treatment cells) - ΔCT (control),

where Ct was the threshold cycle value for the amplified genes (Livak and Schmittgen, 2001), then the fold changes were measured as $2^{-\Delta\Delta CT}$.

Statistical analysis

The average genes expression in the treated cells with lactoferrin as well as the control cells were compared using the Tukey test in One-way ANOVA using SPSS V. 21.

Results

The purity of lactoferrin was confirmed on 10% SDS-PAGE gel electrophoresis and the 80 kDa band of lactoferrin observed on the gel (Figure 1). The concentration of lactoferrin was determined as 3 mg/mL.

Cell viability using MTT assay

To evaluate the cytotoxic activity of lactoferrin against human breast cancer cells (MCF-7) the cell survival was measured using MTT assay. The results showed that the viability rates of the cells in treatments with lactoferrin concentrations of 0, 50, 300, 600, and 800 µg/mL after 24 hours were 100%, 94%, 83%, 62%, and 32%, respectively (Figure 2). By increasing the concentration of lactoferrin, the viability of the cell decreases, as a result, the effect of lactoferrin is dose-dependent.

cDNA synthesis and PCR reaction

After the synthesis of cDNA, the polymerase chain reaction with specific primers was done for three genes. The bands of amplified genes were observed on the agarose gel (Figure 3).

Study of gene expression by real-time PCR

Gene expression levels were evaluated after treatment of three concentrations of 0, 50 and 800 µg/mL of lactoferrin for 24 h on MCF7 cells. The genes amplification curves are visible in the following graph (Figure 4). The fold change in gene expression determined based on $\Delta\Delta CT$ and the amount of $-\Delta\Delta CT$ and $2^{-\Delta\Delta CT}$ are presented in Tables 2 and 3.

The expression level of the Bax gene at concentrations of 50 µg of lactoferrin increased by 2.71 fold and in 800 µg decreased by 0.88 fold. The expression level of the Bak gene at a concentration of 50 and 800 µg of lactoferrin increased by 1.23 and 1.01 fold, respectively. Statistical analysis of the data indicated that the expression levels of two genes of *Bax* and *Bak* at a concentration of 50 µg/mL of lactoferrin were significantly increased, compared with the control

Table 1. Primers used for amplification of the genes

Gene	Sequence (5'-3')	Product Size (bp)
β -actin/F β -actin/R	CCTGGGCATGGAGTCCTGT ATCTCCTTCTGCATCCTGTCG	151 bp
Bax/F Bax/R	CGAGTGTCTCAAGCGCATC GGTCTGATCAGTCCGGCAC	150 bp
Bak/F Bak/R	GCTCGCCATCATCGGGGAC CAACAGGCTGGTGGCAATC	127 bp

**Table 2.** The amount of $-\Delta\Delta CT$ and $2^{-\Delta\Delta CT}$ for Bak

$2^{-\Delta\Delta CT}$	$\Delta\Delta CT$	ΔCT	Treatment
1	0	6.85	0
1.23	- 0.03	6.55	50
1.01	- 0.01	6.86	800

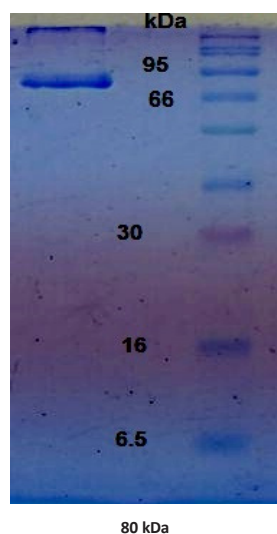
**Table 3.** The amount of $-\Delta\Delta CT$ and $2^{-\Delta\Delta CT}$ for Bax

Treatment	ΔCT	$\Delta\Delta CT$	$2^{-\Delta\Delta CT}$
0	7.06	0	1
50	5.62	- 1.44	2.71
800	7.25	0.19	0.88

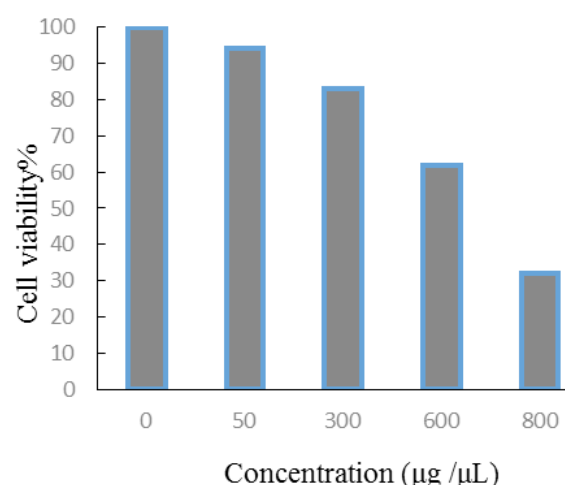


($P < 0.01$). At a concentration of 800 $\mu\text{g/mL}$ of lactoferrin, the expression of the Bak gene increased compared with the control and was statistically significant; however, the expression of the Bax gene was significantly lower than the control gene ($P < 0.02$). According to the results of *Bax* and

Bak genes expression at the concentration of 50 $\mu\text{g/mL}$ of lactoferrin, expression increased, which is likely to be more effective in expressing these genes in this type of cancer (Figure 5). For each sample, measurements were done at least in triplicates; error bars represent standard deviations.

**Figure 1.** 10% SDS-PAGE

Line 1, BLUtra Prestained Protein Ladder (GeneDrex); line 2, 80 kDa band of purified lactoferrin

**Figure 2.** The percentage of viability of breast cancer cells after treatment with different concentrations of lactoferrin

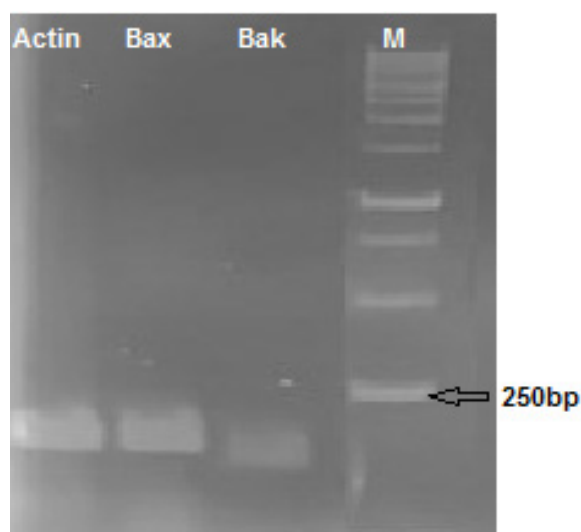


Figure 3. Agarose gel of the amplified genes

Line 1, 151 bp band of β -actin; line 2, 150 bp band of *Bax* gene; Line 3, 127 bp band of *Bak* gene; Line 4, 1kbDNA ladder, Promega.

According to the results of this study and previous studies, the effect of lactoferrin on breast cancer cells can be described schematically (Figure 6). On the extrinsic pathway, it can increase the activity of caspase 3 and induce apoptosis. In the intrinsic pathway with the direct effect on caspase 9, as well as an indirect effect on the *Bax* and *Bak* complex, and their attachment to mitochondrial membrane and release of cytochrome C induce apoptosis. In this pathway, cytoplasmic cytochrome-C transiently binds the caspase adaptor molecule, Apaf-1, and consequently binds to procaspase 9 to form the apoptosome. In this complex structure, procaspase-9 dimerizes and auto-activates. Activated caspase-9 then

stimulates the caspase 3 in the extrinsic pathway, thus inducing apoptosis.

Discussion

Lactoferrin can inhibit the growth of cancer cells. In the last decade, lactoferrin has been given special attention to using as a portion of healthy natural food (milk), due to its ability to act as an anti-cancer therapeutic. In the present study, lactoferrin in a concentration of 50 to 800 $\mu\text{g}/\text{mL}$ could inhibit the growth of breast cancer cells MCF7 in a dose-dependent manner. The effects of bovine lactoferrin on human breast cancer HS578T and T47D cells showed that lactoferrin concentration from 0.125 to 125

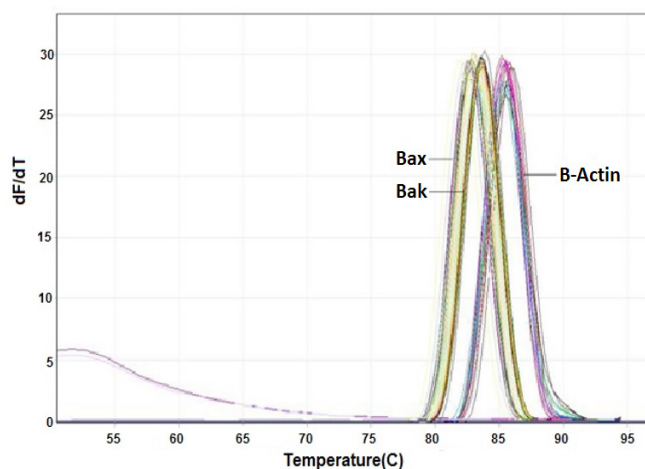


Figure 4. The graph of the proliferation of β -actin, *Bax* and *Bak* genes in a real-time PCR

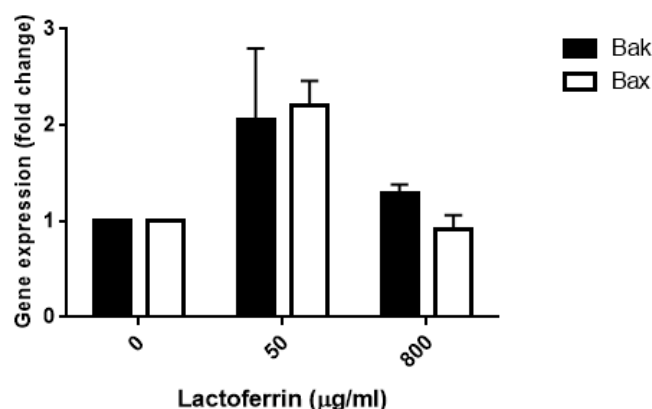


Figure 5. Comparison of the changes in the expression of Bak and Bax genes in different concentration of lactoferrin

Columns represent the fold change of *Bax* and *Bak* genes after treatment with lactoferrin relative to untreated cells. For each sample measurement were done in triplicates, error bars represent SDs ($P < 0.02$).

µM decreased the cell viability of HS578T and T47D by 47% and 54%, respectively [14]. Bovine lactoferrin with a concentration of 500 µg/mL inhibited the gastric cancer cells AGS growth by 55% and 80% during 20 and 48 hours, respectively [7]. The concentration of 500 µg/mL of lactoferrin reduced cell viability in esophageal cancer cell line KYSE by 53% and 80% after 20 and 62 hours, respectively [8]. Lactoferrin B (200 µg/mL) was

cytotoxic for different human leukemia and carcinoma cell lines [24]. Bovine lactoferrin significantly inhibited colon, esophagus, lung, and bladder carcinogenesis in rats when administered orally in the post-initiation stage [6]. The concentration of 30 µM of bovine lactoferrin inhibits the growth of breast cancer cells such as MDA-MB-231, HS578T, and MCF7 [29]. Both bovine lactoferrin and LferricinBs considerably reduced the viability

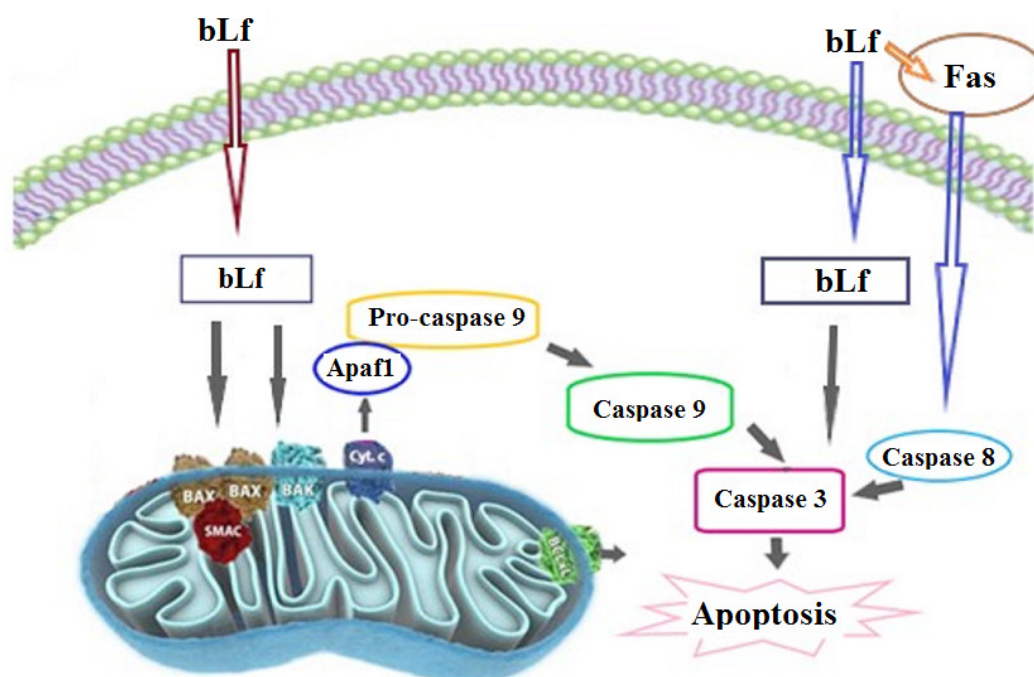


Figure 6. Lactoferrin enters the cell through its receptor and then activates the two pathways

One pathway by activating the caspases (extrinsic) and another through the mitochondrial pathway (intrinsic)

of colon cancer cells HT-29 after 12 h of treatment and significantly induced apoptosis in these cells [30].

Lferricin can activate the intrinsic pathway of apoptosis by the induction of caspases 2, 3, and 9 activities in Jurkat T leukemia cells and MDA-MB-435 breast carcinoma cells [24]. Since caspases 2 and 9 act as initiating caspases for mitochondrial-dependent apoptosis, their involvement in Lfcin-induced apoptosis plays an important role in the mitochondria-dependent pathway of apoptosis and the destruction of human cancer cells [24]. Distribution of mitochondrial membrane potential is usually associated with mitochondrial membrane instability, which results in the release of cytochrome-C and other mitochondrial apoptotic proteins in the cytoplasmic compartment [24].

In this study, lactoferrin at a concentration of 50 µg/mL could increase the expression of *Bax* and *Bak* genes. In the previous study, the expression of the *Bax* gene increased in mice with a tumor that fed with lactoferrin [11]. Lactoferrin inhibits the tumor growth in the colon of the azoxymethane-treated rat, which is related to the apoptosis of these cells induced by the activation of the Fas signaling pathway and enhancing Fas expression [11]. Apoptosis is a highly regulated process controlled by several signaling pathways, such as caspases and mitochondrial pathways [31].

It is known that lactoferrin induces apoptosis by the extrinsic pathway (Fas receptor family and caspases) and intrinsic pathway (mitochondria-associated). The process depends on the cell type [29]. *Bax* and *Bak* the members of Bcl-2 family are the two key molecules in the mitochondrial pathway of apoptosis. These pro-apoptotic Bcl-2 proteins after activation, change their conformation and oligomerize into clusters at the outer membrane of mitochondria resulting in the release of cytochrome-C which may indirectly activate caspase 3 from caspase 8, 9 or directly by caspase 9 [21].

In the present study, the effect of lactoferrin at a concentration of 800 µg, showed an increase in *Bax* gene expression but *Bak* gene expression decreased in comparison to the control gene. This fact indicates that cancer cells have a very contradictory and unpredictable behavior about tumor suppressor and oncogenes genes. Also, many cancer cells inhibit the expression of tumor suppressor genes to increase their rate of growth. Furthermore, cancer cells can escape apoptosis, and therefore apoptosis promotion is one of the mechanisms to fight cancer [32]. Therefore, studying the effect of reagents on increasing the expression of genes involved

in the pathway of apoptosis is important. *Bax* and *Bak* play a crucial role in the regulation of apoptosis. Heterodimerization between Bcl-2, *Bax*, and *Bak* mediated through BH3 domains is removed in many cancers [33]. Therefore, lactoferrin can induce this complex thereby facilitating the induction of apoptosis in cancerous cells. Activation of the pro-apoptotic proteins such as *Bax* and *Bak* is created through several mechanisms depending on each other and involve their translocation from the cytosol to the mitochondria membrane following the induction of apoptosis [34]. The expression of *Bax* and *Bak* genes can be influenced by other genes of the Bcl-2 family, cancer type, and different concentrations of lactoferrin. As a result, lactoferrin alone and directly cannot affect the activity and amount of these proteins. Considering the role and mechanism of lactoferrin in the pathway of apoptosis as inducing factors, we can expect the difference in the expression of genes such as *Bax* and *Bak* involved in this pathway.

Conclusion

The effect of lactoferrin on reducing the survival of cancer cells is dose-dependent and by increasing the concentration of lactoferrin, the viability of the cells decreases. According to the results, *Bax* and *Bak* genes expression increased at the concentration of 50 µg/mL of lactoferrin, which is likely to be more effective in expressing these genes in this type of cancer and may be considered as a defense anti-cancer mechanism.

Ethical Considerations

Compliance with ethical guidelines

This article does not contain any studies with human participants or animals performed by any of the authors. In this study, informed consent was obtained from all individual participants included.

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Authors contribution's

Conceptualization and supervision: Fatemeh Moradian and Alireza Rafiei; Matodology: Amir Khalafi; Investigation, writing and review: All authors; Writing the first draft: Amir Khalafi; Performed the review: Fatemeh Moradian; Commented on the last version; funding ac-

quisition resources: Alireza Rafiei, Fatemeh Moradian; and all authors read and approved the final manuscript.

Conflict of interest

The authors declared no conflict of interest.

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