

Synthesis and Characterization of Nanostructured Lipid Carriers Containing Tretinoin and Grape Seed Oil Using Hot Homogenization and Ultrasonic Waves





Rokhsaneh Balouchi¹, Mahdie Arefi², Sara Daneshmand^{1*}

- 1. Department of Pharmaceutics, Faculty of Pharmacy, Zabol University of Medical Sciences, Zabol, Iran.
- 2. Medical Research Center, Zabol University of Medical Science, Zabol, Iran.



Citation Balouchi R, Arefi M, Daneshmand S. Synthesis and Characterization of Nanostructured Lipid Carriers Containing Tretinoin and Grape Seed Oil Using Hot Homogenization and Ultrasonic Waves. Research in Molecular Medicine. 2024; 12(1):49-56. https://doi.org/10.32598/rmm.11.4.1380.1



doj* https://doi.org/10.32598/rmm.11.4.1380.1

Article Type:

Research Paper

Article info:

Received: 10 Sep 2023 Revised: 23 Sep 2023 Accepted: 05 Dec 2023

Keywords:

Nanostructured lipid carriers (NLCs), tretinoin, Formulation optimization, hot homogenization, grape seed oil (GSO)

ABSTRACT

Background: This study focuses on the formulation and characterization of nanostructured lipidcarriers (NLCs) containing tretinoin, a vitamin A derivative known for its anti-aging and antiacne properties. The primary objective was to enhance drug loading capacity and reduce skin irritation while prolonging the duration of action.

Materials and Methods: Utilizing hot homogenization and ultrasound methods, 26 different formulations were developed, varying lipid and surfactant percentages, as well as ratios of Tween to Span. The optimized NLCs were assessed for morphology, particle size, and zeta potential, revealing that increasing lipid and tretinoin concentrations led to larger sizes and a consistent zeta potential. Morphological analysis confirmed the spherical nature of the nanoparticles, and FTIR spectroscopy indicated successful drug encapsulation.

Results: The optimal formulation comprised 2% lipid, 2% surfactant, and 0.01% tretinoin, achieving a particle size of 253.4 nm and a polydispersity index (PDI) of 0.241.

Conclusion: Overall, these results underscore the promise of NLCs in advancing topical drug delivery systems, paving the way for future clinical applications and further research into their efficacy and safety in diverse dermatological conditions.

* Corresponding Author:

Sara Daneshmand, Assistant Professor.

Address: Department of Pharmaceutics, Faculty of Pharmacy, Zabol University of Medical Sciences, Zabol, Iran.

Phone: +98 (616) 15881 E-mail: sdmehrpooya@gmail.com





Introduction

he synthesis and characterization of lipid carriers at the nanoscale have garnered significant interest in drug delivery and cosmetic formulations, particularly through the development of nanostructured lipid carriers (NLCs). These systems combine the advantages of solid lipid nanoparticles (SLNs) and liquid lipid formulations, representing a significant advancement over SLNs. NLCs offer several advantages, including higher drug loading capacity, reduced drug expulsion, improved drug release profiles, enhanced stability, and superior tolerance to processing conditions [1]. Tretinoin (all-trans retinoic acid) is well-known for its effectiveness in addressing several skin issues, such as acne, photoaging, and hyperpigmentation. However, its therapeutic potential is often compromised by its limited stability and low solubility in water, which can reduce its effectiveness in topical treatments. By encapsulating tretinoin in lipid carriers, especially NLCs, its stability and solubility can be improved, leading to better therapeutic results. The lipid matrix in NLCs not only safeguards the active compound from degradation but also promotes controlled release, resulting in prolonged therapeutic effects [2, 3].

Grape seed oil (GSO) is an appealing lipid phase for NLC formulations due to its rich composition of polyunsaturated fatty acids and bioactive compounds, such as proanthocyanidins and tocopherols. These components provide antioxidant properties, which can synergistically enhance the stability of tretinoin within the NLCs. The incorporation of GSO not only serves as a lipid carrier but also contributes to the overall efficacy of the formulation, making it particularly suitable for dermatological applications [4-7].

The aim of this study was to encapsulate tretinoin in NLC using hot homogenization followed by ultrasonic treatment. The particle size of the NLC systems was analyzed. Additionally, the encapsulation efficiency (%EE) of the NLC was examined, and fourier transform infrared spectroscopy (FTIR) was employed. This provided insights into potential strategies for enhancing the stability and bioavailability of tretinoin for future use in cosmetic formulations. Moreover, the optimization of the formulation was achieved using response surface methodology (RSM), a statistical approach that facilitates the evaluation of multiple variables and their interactions, ensuring a robust and effective NLC formulation [8]. This comprehensive study underscores the potential of NLCs as a promising vehicle for tretinoin delivery, paving the way for advanced therapeutic solutions in skin care.

Materials and Methods

Materials

Tween 20, Sorbitan monostearate (Span), and stearic acid were obtained from Merck Chemical Co. (Darmstadt, Germany). Tretinoin and GSO were purchased from Sepidaj Pharmaceutical Company, Iran, and Ferdows Juice Manufacturing Company, respectively.

Preparation of NLCs

The synthesis of NLCs was conducted using the hot homogenization method combined with ultrasound waves. Briefly, tretinoin (C₂₀H₂₈O₂, MW= 300.442 g.mol⁻¹, and melting point=180 °C) was dissolved in liquid lipid (GSO), and the mixture was added to the melted solid lipid (stearic acid). The lipid phase was then heated to 70 °C (the water bath set point, which was 5 °C higher than the melting point of the solid lipid). Next, the hot aqueous surfactant solution containing different concentrations of water and Tween (Table 1), with the same temperature as the melted lipid mixture, was added dropwise to the lipid phase under homogenization (Silent Crusher M, Heidolph, Nuremberg, Germany) at 20000 rpm for 45 minutes, followed by exposure to ultrasonic waves for an additional 3 minutes. The hot o/w nanoemulsion was cooled down to room temperature, resulting in lipid phase recrystallization and the ultimate formation of NLCs [9].

Characterization of NLCs

Particle size, zeta potential, and polydispersity index (PDI) analysis

The particle size, zeta potential, and PDI of RG-NLC were assessed using photon correlation spectroscopy (PCS) using a Zetasizer (Malvern, UK) at 25 °C, employing disposable plain folded capillaries. Before the measurements, all samples were diluted with distilled water and vortexed for 30 seconds to achieve an appropriate scattering intensity [10].

Morphological examination

The morphology of the NLCs was assessed using scanning electron microscopy (SEM). Samples were prepared by placing them onto a carbon tape and subjecting them to gold-palladium coating under vacuum. Images were captured to analyze the surface characteristics and shape of the nanoparticles [11].



Table 1. Formulations containing various percentages of stearic acid, tretinoin, and surfactant

No.	Lipid (%W/W)	Tretinoin (%W/W)	Ratio of Lipid	Ratio of Surfactant (Tween/Span)	Surfactant (%W/W)	Size	Zeta Potential	PDI
1	2	0.03	4	1.25	2	325.27.42	-22.33.21	0.3150.081
2	3	0.05	4	0.50	1	549.97.3	-23.83.92	0.5910.073
3	2	0.03	3	1.25	1	382.34.32	-22.24.81	0.3650.041
4	1	0.01	4	2	3	2314.81	-23.95.35	0.2820.047
5	1	0.05	2	2	3	262.33.95	-23.24.85	0.3570.058
6	1	0.01	2	0.5	1	252.37.49	-24.36.83	0.2900.053
7	3	0.03	3	1.25	2	503.96.82	-25.23.20	0.5620.087
8	1	0.05	4	0.5	3	273.26.10	-24.36.31	0.2930.052
9	1	0.03	3	1.25	2	258.35.61	-23.33.55	0.2400.064
10	2	0.03	3	1.25	3	251.23	-24.25.66	0.3390.045
11	2	0.03	3	1.25	2	331.23.11	-22.53.20	0.3310.073
12	3	0.05	2	2	1	541.13.95	-24.37.38	0.5630.068
13	2	0.01	3	1.25	2	253.45.38	-25.23.20	0.2410.071
14	1	0.05	4	2	1	262.39.36	-22.88.38	0.2530.093
15	2	0.03	3	1.25	2	338.34.94	-24.93.92	0.3100.04
16	2	0.03	3	2	2	315.28.22	-22.36.75	0.3220.059
17	3	0.01	4	0.5	3	493.53.81	-23.47.90	0.5450.064
18	3	0.05	2	0.5	3	521.87.32	-25.34.25	0.5830.038
19	2	0.01	3	1.25	2	269.34.88	-24.26.94	0.3010.060
20	2	0.03	3	0.5	2	322.35.30	-23.93.54	0.3980.075
21	2	0.05	3	1.25	2	3534.32	-21.26.43	0.4520.069
22	2	0.03	3	1.25	2	325.35.90	-23.35.48	0.4080.031
23	2	0.03	2	1.25	2	319.24.35	-22.87.35	0.411.058
24	2	0.03	3	1.25	2	302.54.85	-24.36.92	0.4150.062
25	3	0.01	4	2	1	501.93.84	-22.34.92	0.5820.035
26	3	0.01	2	2	3	495.26.31	-25.26.51	0.5020.62

Drug loading efficiency

The drug loading capacity of the NLCs was measured using an indirect method. To assess the amount of tretinoin entrapped in the NLCs, a certain volume of NLC suspension was centrifuged at 6000 rpm for 30 minutes using Amicon filter devices with a 10 kDa cutoff. Sub-

sequently, the supernatant was analyzed to determine the concentration of unencapsulated tretinoin using UV-Vis spectroscopy. Calibration curves were generated using various concentrations of tretinoin dissolved in DMSO to facilitate quantification at a wavelength of 340 nm [12, 13].

ERMM



FTIR analysis

The infrared spectra were recorded using an FTIR spectrophotometer (IRAffinity-1S, Shimadzu, Tokyo, Japan) with a sample-to-KBr ratio of 1:10, at a resolution of 4 cm⁻¹. Scans were conducted within the frequency range of 4000 to 500 cm⁻¹, with one scan performed for each individual outcome [14].

Response surface methodology (RSM)

RSM was employed to optimize the formulation parameters of NLCs containing tretinoin and GSO. The methodology involved the following steps: A central composite design (CCD) was used to systematically vary the independent variables, which included the concentration of lipid, tretinoin, surfactant, and the ratio of surfactant. The design matrix was generated using Design Expert software version 13.0.5.0, resulting in a set of experimental runs to evaluate the effects of these parameters on the responses. Each experimental run was conducted according to the design matrix, and data were collected on the following responses: Particle size, zeta potential, and PDI. The collected data were analyzed using Design Expert software to fit a second-order polynomial model, which describes the relationship between the independent variables and the responses. The software was utilized to determine the optimal formulation conditions that maximize %EE while minimizing particle size.

Statistical analysis

All the experiments were carried out in triplicate and represented as the Mean±SD. Data were analyzed using the Design Expert software, which employs RSM to optimize the formulation parameters and predict interactions between independent variables and their effects on responses [15].

Results

Preparation and optimization of NLCs

Design of experiments

The NLC formulations were synthesized based on a systematic design using the RSM with a total of 26 different formulations. The primary variables included percentages of lipid, surfactants (tween and span), and tretinoin. The specific formulations involved variations, seeking to optimize the %EE and stability of the NLCs. Table 1, designed using RSM software, comprises five

columns that represent the initial data for formulating the desired combination. The DLS data were compared with the predicted data from the software, leading to the identification of formulation number 13 as the optimal formulation by RSM. This formulation contains 0.01% Tretinoin, 2% surfactant, a stearic acid to GSO ratio of 3%, and a Tween to Span ratio of 1.25. The particle characterization, including IR spectroscopy and morphology evaluation using electron microscopy, was performed on the optimal formulation. In the optimal formulation, the size, PDI, and zeta potential were 253.4 nm, 0.241, and -25.2 mV, respectively, while the predicted values for size, PDI, and zeta potential from the software were 247.8 nm, 0.230, and -24.8 mV, respectively.

Error percentage calculation

The error percentages were calculated based on the Equation 1:

1.Error percentage=
$$\frac{\text{DLS data-predicted data} \times 100}{\text{DLS data}}$$

This analysis demonstrated the reliability of the RSM predictions in formulating the desired product, with acceptable error margins across the measured parameters (Table 2).

Characterization of NLCs

Particle size analysis

The sizes of the NLCs were determined using a particle size analyzer. The results for the optimized formula are summarized in Table 1. Notably, the size and PDI showed a notable variation with changing concentration levels of lipids and surfactants. For example, a formulation containing 2% lipid and a surfactant ratio of 1.25:1 produced a particle size of 253.4±5.38 nm with a PDI of 0.241±0.064. The DLS data revealed that as the lipid concentration increased, the particle size also increased, with the optimal formulation being characterized as follows:

Zeta potential

The zeta potential, an important indicator of the stability of nanoparticle dispersions, was measured for each formulation. The optimal formulation exhibited a zeta potential of -25.2 mV, indicating sufficient stability for pharmaceutical use. According to the literature, a zeta potential threshold of ± 30 mV is commonly indicative of good stability; hence, the observed value supports the potential clinical applicability of the developed NLCs.



Table 2. Error percentages of the data

Variables	DLS Data	Predicted Data	Error (%)
Size	253.4	247.8	2.20
PDI	0.241	0.230	4.5
Zeta potential	-25.2	-24.8	1.5



PDI

The PDI values were analyzed to determine the homogeneity of the formulations. All formulations were within the acceptable range (0-1), with the optimal formulation showing a PDI of 0.241. A lower PDI value suggests a uniform distribution of particle sizes, which is desirable for consistent drug delivery.

Morphological characterization

The morphology of the NLCs was assessed through SEM. The images captured (Figure 1) illustrated that the NLCs had a spherical shape, which is beneficial for penetration into the skin's stratum corneum and enhances the potential for controlled release (Figure 1).

Drug loading efficiency

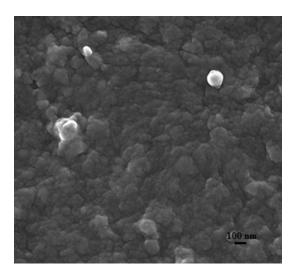
To evaluate the drug loading efficiency, a calibration curve was constructed by measuring the absorbance of various concentrations of tretinoin in DMSO using UV-Vis spectroscopy. The proportion of encapsulated tretinoin was calculated based on the concentration present in the supernatant after centrifugation. The results indicated an %EE of approximately 90%, suggesting that a significant proportion of the drug was successfully loaded into the NLCs.

FTIR spectroscopy

FTIR was employed to confirm the incorporation of tretinoin in the NLC matrix. The FTIR spectra of both the pure tretinoin and the NLC formulation were compared, revealing characteristic peaks of the drug in the NLC spectrum, which suggests successful encapsulation. The spectra showed that the characteristic functional groups of tretinoin were preserved after formulation, although some peaks were diminished due to interactions between the drug and the lipid matrix (Figure 2).

Discussion

The treatment of acne, a chronic inflammatory disorder that predominantly affects adolescents and young adults, requires effective and targeted therapeutic interventions [16]. Acne is characterized by the formation of comedones, papules, pustules, and sometimes scarring,



ERMM

Figure 1. SEM image of the NLCs encapsulating tretinoin

The image illustrates the morphology and uniformity of the NLCs, highlighting their spherical shape and size distribution, which are conducive to enhanced skin penetration and drug delivery efficiency.



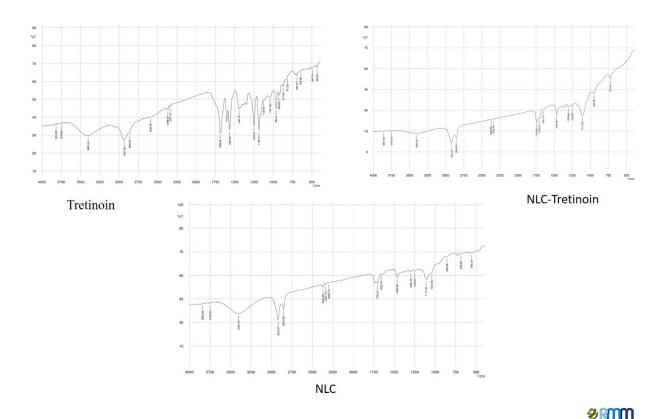


Figure 2. FTIR analysis of tretinoin Tretinoin-NLC, and NLC, illustrating the characteristic absorption bands and chemical interactions among the compounds.

primarily due to the inflammation of pilosebaceous units in the skin [17]. Recent statistics reveal that approximately 85% of individuals between the ages of 12 and 25 experience some form of acne, although it can persist well into adulthood [18]. The impact of acne transcends physical symptoms; it can lead to significant psychological distress, including anxiety, depression, and low self-esteem. Notably, managing acne effectively is vital not only for physical health but also for emotional wellbeing [19]. Among various treatment modalities, topical retinoids—particularly tretinoin, the active metabolite of vitamin A—have emerged as one of the most effective options for managing acne vulgaris [20]. Tretinoin exerts its anti-acne action through several mechanisms, including normalization of keratinization, reduction of sebum production, and anti-inflammatory properties. However, despite its therapeutic benefits, tretinoin's clinical use is often hampered by its limited skin penetration, high irritation potential, and instability. These challenges necessitate innovative approaches, such as formulation in nanosized delivery systems, which can enhance the bioavailability, stability, and overall efficacy of the drug while minimizing potential side effects [21]. In response to these challenges, the development of NLCs presents a promising solution. NLCs combine solid lipids and liquid lipids to improve the %EE of lipophilic drugs, such as tretinoin, and facilitate controlled release. Additionally, NLCs can protect the active ingredient from environmental degradation and improve overall stability, which is crucial for maintaining therapeutic efficacy [22, 23].

The primary objective of this research was to design and synthesize NLCs containing tretinoin and GSO using a combination of hot homogenization and ultrasonication techniques. The incorporation of GSO as a lipid phase in NLC formulations adds another layer of efficacy. GSO, rich in linoleic acid and antioxidants, was selected for this study due to its beneficial properties for skin health, including its moisturizing effects and potential anti-inflammatory activity, which can complement the action of tretinoin [24, 25].

A previous study has shown that oils rich in linoleic acid, like GSO, can improve skin hydration and may possess anti-inflammatory properties, further supporting its use in formulations aimed at acne treatment [26]. Therefore, in this study, GSO was used as the liquid lipid in the formulation. We utilized hot homogenization followed by ultrasonic method to encapsulate tretinoin in NLCs, building on findings from a 2013 study by Nasrollahi et al., which indicated that the use of a high-pres-



sure homogenizer to produce SLNs of tretinoin resulted in a formulation with a slower and longer-lasting effect compared to tretinoin cream, suggesting that the SLN formulation has better skin tolerance and could serve as a more effective option in topical treatments [27]. The resulting formulations were characterized for particle size and %EE. The morphological analysis via SEM revealed spherical nanoparticles, which is beneficial for ensuring uniform penetration across the skin barrier [28]. Additionally, FTIR was employed to analyze the interactions between the lipid matrix and the encapsulated drug. FTIR analysis revealed characteristic peaks associated with the presence of tretinoin within the lipid matrices, suggesting successful encapsulation within the NLCs. The FTIR spectrum of pure tretinoin exhibited several significant peaks corresponding to specific functional groups in its molecular structure. Key peaks included a peak at 2931 cm⁻¹ attributed to the stretching vibrations of the (-C-H) group, peaks at 1188 cm⁻¹ and 1249 cm⁻¹ corresponding to the stretching vibrations of the (C-O) group, and a peak at 1689 cm⁻¹ associated with the stretching vibrations of the (C=O) bond.

Additionally, the trans configuration of the vinyl group was represented by a peak at 1964 cm⁻¹, while the stretching vibrations of (C=C) coupled with the carbonyl group were observed at 1566 cm⁻¹ with high intensity. As illustrated, the FTIR spectrum of pure tretinoin displays sharp and intense peaks, indicating the presence of various functional groups. Furthermore, the FTIR results for the blank sample and the NLC formulation containing tretinoin showed striking similarities, with notable reductions in peak intensity for the functional groups in the NLC formulation. This reduction suggests successful encapsulation of tretinoin within the nanoparticles. Importantly, peaks associated with functional groups were absent in formulations lacking the active pharmaceutical ingredient, further confirming the effective incorporation of tretinoin into the NLC. Overall, these results indicate not only successful encapsulation of tretinoin but also optimal particle sizes that enhance skin penetration, thereby supporting the potential efficacy of the NLC formulation in topical applications.

Conclusion

The research successfully developed and characterized NLCs for the effective delivery of tretinoin. The optimized formulation demonstrated a suitable particle size, zeta potential, and %EE, indicating its potential for enhanced skin permeation and reduced irritation compared to traditional formulations. The morphological analysis confirmed the uniformity and stability of the nanopar-

ticles, highlighting their suitability for topical applications. Future studies should focus on in vivo evaluations to assess the pharmacokinetics and therapeutic efficacy of the optimized NLC formulation in acne treatment. Additionally, exploring the incorporation of other active ingredients within the NLC system could enhance multifunctional properties, targeting not only acne but also signs of aging. Investigating alternative methods for large-scale production and stability studies under various storage conditions will also be crucial for the eventual commercialization of this formulation. Finally, patient-centric clinical trials will be essential to validate the safety and effectiveness of the NLCs in real-world applications.

Ethical Considerations

Compliance with ethical guidelines

This study was approved by the Ethics Committee of Zabol University of Medical Sciences, Zabol, Iran (Code: IR.ZBMU.REC.1400.005).

Funding

This study was financially supported by Zabol University of Medical Sciences, Zabol, Iran.

Authors contribution's

Data collection and data analysis: Rokhsaneh Balouchi; writing the original draft: Mahdie Arefi; Conceptualization, supervision and validation: Sara Daneshmand; Final approval: All authors.

Conflict of interest

The authors declared no conflict of interest.

References

- [1] Shirodkar RK, Kumar L, Mutalik S, Lewis S. Solid lipid nanoparticles and nanostructured lipid carriers: Emerging lipid based drug delivery systems. Pharm Chem J. 2019; 53(5):440-53. [DOI:10.1007/s11094-019-02017-9]
- [2] Ghasemiyeh P, Moradishooli F, Daneshamouz S, Heidari R, Niroumand U, Mohammadi-Samani S. Optimization, characterization, and follicular targeting assessment of tretinoin and bicalutamide loaded niosomes. Sci Rep. 2023; 13(1):20023. [DOI:10.1038/s41598-023-47302-6] [PMID]



- [3] Kontzias C, Zaino M, Feldman SR. Tretinoin 0.1% and benzoyl peroxide 3% cream for the treatment of facial acne vulgaris. Ann Pharmacother. 2023; 57(9):1088-93. [DOI:10.1177/10600280221147338] [PMID]
- [4] Yarovaya L, Waranuch N, Wisuitiprot W, Khunkitti W. Chemical and mechanical accelerated and long-term stability evaluation of sunscreen formulation containing grape seed extract. J Cosmet Dermatol. 2022; 21(11):6400-13. [DOI:10.1111/jocd.15308] [PMID]
- [5] Krasodomska O, Paolicelli P, Cesa S, Casadei MA, Jungnickel C. Protection and viability of fruit seeds oils by nanostructured lipid carrier (NLC) nanosuspensions. J Colloid Interface Sci. 2016; 479:25-33. [DOI:10.1016/j.jcis.2016.06.041] [PMID]
- [6] Turcu RP, Panaite TD, Untea AE, Vlaicu PA, Badea IA, Mironeasa S. Effects of grape seed oil supplementation to broilers diets on growth performance, meat fatty acids, health lipid indices and lipid oxidation parameters. Agriculture. 2021; 11:404. [DOI:10.3390/agriculture11050404]
- [7] Nakamura Y, Tsuji S, Tonogai Y. Analysis of proanthocyanidins in grape seed extracts, health foods and grape seed oils. J Health Sci. 2003; 49(1):45-54. [DOI:10.1248/jhs.49.45]
- [8] Chen WH, Uribe MC, Kwon EE, Lin KY, Park YK, Ding L, et al. A comprehensive review of thermoelectric generation optimization by statistical approach: Taguchi method, analysis of variance (ANOVA), and response surface methodology (RSM). Renew Sustain Energy Rev. 2022; 169:112917. [DOI:10.1016/j.rser.2022.112917]
- [9] Mohammadi M, Pezeshki A, Mesgari Abbasi M, Ghanbarzadeh B, Hamishehkar H. Vitamin D3-loaded nanostructured lipid carriers as a potential approach for fortifying food beverages; in vitro and in vivo evaluation. Adv Pharm Bull. 2017; 7(1):61-71. [DOI:10.15171/apb.2017.008] [PMID]
- [10] Ebrahimian M, Shahgordi S, Yazdian-Robati R, Etemad L, Hashemi M, Salmasi Z. Targeted delivery of galbanic acid to colon cancer cells by PLGA nanoparticles incorporated into human mesenchymal stem cells. Avicenna J Phytomed. 2022; 12(3):295-308. [PMID]
- [11] Daneshmand S, Yazdian-Robati R, Jaafari MR, Movaffagh J, Malaekeh-Nikouei B, Iranshahi M, et al. Evaluation of the anti-melanogenic activity of nanostructured lipid carriers containing auraptene: A natural anti-oxidant agent. Nanomed J. 2022; 9(1):57-66. [DOI:10.22038/nmj.2022.62354.1645]
- [12] Madane RG, Mahajan HS. Curcumin-loaded nanostructured lipid carriers (NLCs) for nasal administration: Design, characterization, and in vivo study. Drug Deliv. 2016; 23(4):1326-34. [DOI:10.3109/10717544.2014.975382] [PMID]
- [13] Sarkis N, Sawan A. Development and validation of derivative UV spectroscopic method for simultaneous estimation of nicotinamide and tretinoin in their binary mixtures and pharmaceutical preparations. BMC Chem. 2022; 16(1):15. [DOI:10.1186/s13065-022-00809-x] [PMID]
- [14] Saeedi M, Rezanejad Gatabib Z, Morteza-Semnani K, Rahimnia SM, Yazdian-Robati R, Hashemi SMH. Preparation of arbutin hydrogel formulation as green skin lightener formulation: In vitro and in vivo evaluation. J Dispers Sci Technol. 2024. [DOI:10.1080/01932691.2024.2371954]
- [15] Shiehzadeh F, Mohebi D, Chavoshian O, Daneshmand S. Formulation, characterization, and optimization of a topical

- gel containing tranexamic acid to prevent superficial bleeding: In vivo and in vitro evaluations. Turk J Pharm Sci. 2023; 20(4):261-9. [DOI:10.4274/tjps.galenos.2022.60687] [PMID]
- [16] Layton AM, Ravenscroft J. Adolescent acne vulgaris: Current and emerging treatments. Lancet Child Adolesc Health. 2023; 7(2):136-44. [DOI:10.1016/S2352-4642(22)00314-5] [PMID]
- [17] Najeeb A, Gaurav V. "Acne" terminology in dermatology. CosmoDerma. 2024; 4:4. [DOI:10.25259/CSDM_244_2023]
- [18] Kostecka M, Kostecka J, Szwed-Gułaga O, Jackowska I, Kostecka-Jarecka J. The impact of common acne on the wellbeing of young people aged 15-35 years and the influence of nutrition knowledge and diet on acne development. Nutrients. 2022; 14(24):5293. [DOI:10.3390/nu14245293] [PMID]
- [19] Sieradocha K. The mental health implications of Acne Vulgaris. Qual Sport. 2024; 35:56063. [DOI:10.12775/ QS.2024.35.56063]
- [20] Eichenfield DZ, Sprague J, Eichenfield LF. Management of acne vulgaris: A review. JAMA. 2021; 326(20):2055-67. [DOI:10.1001/jama.2021.17633] [PMID]
- [21] Zhong J, Zhao N, Song Q, Du Z, Shu P. Topical retinoids: Novel derivatives, nano lipid-based carriers, and combinations to improve chemical instability and skin irritation. J Cosmet Dermatol. 2024; 23(10):3102-15. [DOI:10.1111/ jocd.16415] [PMID]
- [22] Chutoprapat R, Kopongpanich P, Chan LW. A mini-review on solid lipid nanoparticles and nanostructured lipid carriers: Topical delivery of phytochemicals for the treatment of Acne Vulgaris. Molecules. 2022; 27(11):3460. [DOI:10.3390/ molecules27113460] [PMID]
- [23] Safta DA, Bogdan C, Moldovan ML. SLNs and NLCs for skin applications: Enhancing the bioavailability of natural bioactives. Pharmaceutics. 2024; 16(10):1270. [DOI:10.3390/ pharmaceutics16101270] [PMID]
- [24] de Castro MLTP. A new insight of grape seed extract in skincare [MA thesis]. Lisbon: Universidade Catolica Portuguesa; 2024. [Link]
- [25] Tao K, Guo L, Hu X, Fitzgerald C, Rouzard K, Healy J, et al. Encapsulated activated grape seed extract: A novel formulation with anti-aging, skin-brightening, and hydration properties. Cosmetics. 2021; 9:4. [DOI:10.3390/cosmetics9010004]
- [26] Musallam AA, Elmahboub Y, Attalla KZ, Elfarouk O, Aly N, Bligh L, et al. Formulation of grape seed oil and hibiscus extract for treatment of acne: optimization using D-optimal design and in vitro characterization. J Pharm Sci Drug Manuf Misr Univ Sci Technol. 2025; 2(2):72-80. [Link]
- [27] Nasrollahi SA, Abbasian AR, Farboud ES. In vitro comparison of simple tretinoin-cream and cream loaded with tretinoin-SLN. J Pharm Technol Drug Res. 2013; 2(1):13. [DOI:10.7243/2050-120X-2-13]
- [28] Morais RP, Hochheim S, de Oliveira CC, Riegel-Vidotti IC, Marino CEB. Skin interaction, permeation, and toxicity of silica nanoparticles: Challenges and recent therapeutic and cosmetic advances. Int J Pharm. 2022; 614:121439. [DOI:10.1016/j.ijpharm.2021.121439] [PMID]