

Genetic Variations in Host Factors and their Critical Role on HCV Medication

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Abstract

Although standard treatments are nearly life-saving in hepatitis C patients, all patients equal don't respond to such medications. To achieve an efficient therapeutic response in all patients, medications should individually alter based on genetic factor variations. Hence, having more detailed knowledge of the affective factors on the disease promotion could help us to organize an appropriate regime to treat HCV patients. In order to review recent studies on HCV-related treatments, the data was collected from the clinical and molecular studies published during the last 15 years in electronic databases such as PubMed, Google Scholar, Medline, Scopus and Web of Sciences. Thus, this literature reviews the recent clinical studies with the genetic-based aspects, especially those factors affecting the etiopathological and histological feature of hepatitis C, diagnostic genetic and viral affecting criteria as well as some effective treatment strategies.

Keywords: Hepatitis C Virus; Genetic Host Factors; Sustained Virologic Response

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Introduction

Hepatitis C is one of the rising disastrous problems worldwide. Approximately, 170-200 million people 3-3.5% of the world population suffer from hepatitis C virus (HCV) infection with the risk of morbidity and mortality (1-3). Around 3-4 million persons are infected each year and more than 350,000 people die every year from hepatitis C-related liver diseases (4-6).

Amongst different groups of the patients with acute HCV infection, only 15-30% could spontaneous clear the virus while the other patients have potential for promoting severe progressive liver damage (7, 8). Based on the literature reports, about 70-85% of HCV infected patients develop to the chronic stage, and in 20-30% of them it leads to liver cirrhosis associated with hepatocellular carcinoma (HCC) with 1-5% death rate caused by liver cancer (8,9). Considering that several epidemiological studies were conducted in different societies, up to 80% of HCC cases were reported in East Asia and Sub Saharan Africa while only 13% occurred in America (10).

Therefore, from the point of epidemiologic view, HCV infection includes unequal frequencies in different geographic regions; so that, the highest rate of HCV infection occurs in Far East Asian countries (11). Taken together, the prevalence of HCV in some countries of Far East Asia, Africa, South East Asia and East Mediterranean are higher than Europe and North American countries (12, 13, 14).

Hepatitis C virus

HCV is a small, enveloped positive single-stranded ribonucleic (RNA) virus from hepacivirus genus of Flaviviridae family lacking cytopathic effects (15). As illustrated in Figure 1, whole genome of HCV is 9.6 kb which is flanked by conserved non-translated regions (NTRs). These NTR sequences are required for both RNA translation and replication of HCV genes. An internal ribosome site (IRS) is located near the 5'- NTR, conducting the expression of a polyprotein precursor. Core E1 and E2 are structural proteins, originated from precursor polyprotein

cleavage by alternative translation mechanism. Non-structural genes express several proteins, which included P7, NS2, NS3, NS4, NS4B, NS5A and NS5B. Proliferation of the HCV genome occurs when the genome converted to a double-strand RNA and then negative-strand RNA is used as a replicative template (16, 17). Replication process of HCV genome is driven by an RNA-dependent RNA

polymerase, which product of this reaction is a double-stranded RNA. Subsequently, ATP-dependent NS3 helicase converts the double-stranded RNA into positive single-stranded RNA (18, 19). Despite many current advances in HCV proliferation, there is yet no exact mechanism of infection progression (20).

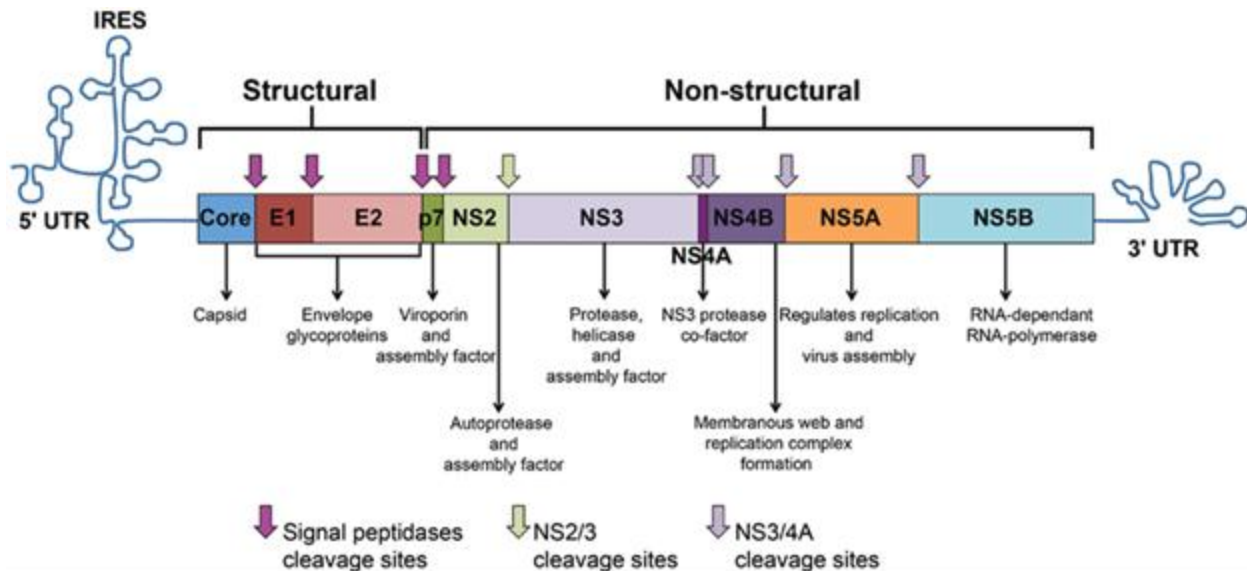


Figure 1. Hepatitis C virus: Model structure and genome organization (21).

The main ways of HCV infection include: 1) Blood transfusion from infected persons, 2) Organ transplant from infected persons; 3) Needlestick cases in hospitals or in health care units; 4) Injection drug users who used sharing syringe; 5) Being born to a hepatitis C-infected mother; 6) Through sex with infected persons; 7) Sharing personal items which were contaminated with infectious blood (4, 22). The HCV infection usually has a different latency period which might take 2 weeks to 6 months; however, about 80% of persons present no symptoms in initial stages of infection (23). On the other hand, some early symptoms including fever, jaundice, abdominal pain, especially pain under the right rib, grey-colored faces, dark urine, vomiting, nausea, fatigue and decreased appetite are the most common indicators for acute infection. However, most patients exhibit no symptoms, which caused a serious difficulty in diagnosing HCV infection (24). The current major approach to detection of HCV infection is the application of anti-HCV, which followed by the RIBA and viral RNA tests. Although many studies performed to design efficient vaccines against HCV, up to now, no one has been able to succeed in such way; hence, it is only treated with antiviral medicines (7).

The most vulnerable people to catch the blood-borne diseases are IV drug users, who are often infected to ADIS and hepatitis, because of collective use of injection equipment and drug solution (25). HCV transmission in 90% of patients were has occurred owing to the use of sharing syringe. Drugs administration and also alcohol abuse are two important co-factors, led to the promotion of disease stage and ultimately, increase mortality and morbidity of HCV patients. Additionally, in most of these patients, at least one co-infection is observed with HIV, EBV and HBV (3). This review focused on viral and host genetic factors possibly affecting the response to treatment, difficulties in HCV clearance and persistence status in patients with hepatitis C.

Viral factors

Treatment of hepatitis C is influenced by the viral factors including viral genotype, quasi-species, baseline viral load and viral kinetics (i.e., rapid and/or early virological response) which can be used as predictors of response (5, 26). Based on molecular analyses, genotype of HCV has been highly diversified by a large number of mutations, which caused wide variety of single nucleotide

polymorphisms (SNPs) in the HCV genome. As such, various genotypes of HCV show 30-35% nucleotide diversity, and at least seven distinct genetic varieties have been identified based on phylogenetic and sequence analyses of complete viral genomes that are

distributed in different geographical regions (27). Therefore, several genotypes of HCV have been identified several in communities worldwide. Considering the extracted data from literature, 6 different genotypes are introduced in Table 1.

Table 1. HCV prevalence in different countries and geographic areas based on genotypic diversities.

Genotype no.	Prevalence location	References
1a	North America and Western Europe	27, 28
1b	United States, Europe and Japan	3
2	developed countries such as	3, 18
3	Thailand and Southeast Asia	18
4	North and Central Africa as well as Middle East countries	27, 28
5	Zimbabwe and other South African countries	18, 27, 28
6	South East Asia such as Hong Kong, Thailand, Indonesia, China, Vietnam, Myanmar and Korea	3, 11, 27, 28, 29

Some mutations in HCV replication emerges from multiple quasi-species of HCV. These emerging genotypes of HCV enable to evade host immune response as well as antiviral drugs through resistance pathways that have been activated by new mutations (18). Therefore, for treating HCV patients, it is a need to identify the type of virus regarding genotypic varieties and single nucleotide polymorphism located in virulence genes of the HCV. The literature has reported that the highest SVR rate is for the patients infected by HCV genotype 2 and the lowest rate in genotypes 1 and 4 (16). Shortening the treatment period and lowering ribavirin dose reduced SVR rate for patients with HCV genotype 1, but not for genotypes 2 and 3. For this reason, some clinical trial studies specifically focused on genotypes 2 and 3 (29, 30).

Additionally, genotype 2 and 6 are more responsive than genotype 1a while, genotype 1b has the highest resistance compared to genotype 2, 6 and 1a (31-33). Although, genotypes 5 and 6 have high similarity to genotypes 3 and 2 respectively, SVR for each genotype differs. Taken together, SVR rate of 40-50% is achieved by following PEG-IFN/RBV therapy in patients with genotype 1, nearly 60% in patients with genotype 4 and 80% in patients with genotype 2 or 3 (5, 13, 34).

Host factors

Applying an efficient treatment against HCV requires a versatile knowledge about host factors as one of the major issues in achieving a higher SVR. The crucial factors affecting HCV-related infections included age, race, gender, obesity, steatosis, disease severity, alcohol intake, host immune status and genetic

factors (26). Numerous researchers have conducted many experiments on the function of host immune system to determine therapy-induced responses when healthy people are infected with HCV (5, 35). These studies have indicated that immune response and antiviral therapy of HCV infection are strongly altered by multiple host genetics factors. Two important factors including cytokines and chemokines are the main players in the promotion of hepatitis C. These factors have been documented to have key roles in the regulation of innate and adaptive immune response to HCV infection (36-38). Multiple studies have found cytokines gene polymorphisms influence on the pathogenesis of HCV and outcome of response (39, 40). In the literature review, the key roles of the most important host factors associated with HCV infection along with all related genes will be discussed.

1. IL-4

One of the major cytokines involved in HCV infection is IL-4, secreted by TH2. Therefore, TH2 is responsible for progression or persistence of HCV infection. High-elevated level of IL-4 suppresses inflammatory cytokines leading to lower liver injury. Besides, this occurrence is associated with inability of host to eliminate the virus and progression of a chronic disease (41). The critical role of genetic varieties in the IL-4 gene has recently been reported. In this relation, the allele C (+33) of the IL-4 gene is observed in acute more than the chronic hepatitis. This polymorphism indicated as IL-4 C/T (+33) found to be associated with HCV spontaneous clearance. The IL-4 (+33) T SNP expresses a high level of IL-4 which increases HCV survival rate and

serious liver damage (8).

2. *IL-8*

IL-8 is a cytokine stimulated by Th17 lymphocyte from contact with HCV NS5A protein for inhibiting the antiviral activity of INF during the progressive stage (42, 43). Besides, production of HCV-induced IL-8 often resulted in induction of COX-2 expression, followed by liver cell injury-related inflammation (44).

3. *IL-10*

IL-10 is classified in class-2 cytokines, which is expressed by TH2. This cytokine is known as an anti-inflammatory factor. Additionally, IL-10 acts via down-regulation of the cytokine gene expression in TH1 and co-stimulatory factors on macrophages. The presence of SNPs located in IL-10 promoter, which alters the IL-10 gene expression can affect several pathways in the immune cells. IL-10 gene expression is strongly associated with SNPs located in regulatory site. One of the most important polymorphisms located in IL-10 gene codon-1082 by which GG genotype results in over-expression of IL-10 protein, causes a low pro-inflammatory response, decreases the liver fibrosis and finally promotes highly spontaneous clearance. This SNP genotype causes down-regulation of IL-10 gene expression and thus weakens antiviral response and contributes HCV infection towards hepatitis chronicity (8).

4. *TH17 Modulatory Cytokines*

T helper 17 (TH17) is known as a T cell belonging to a subgroup of TCD4+ which contributes to autoimmune-related inflammatory diseases with production of immune modulators such as IL-17A, IL-17F or IL-22 and IL-16. These cytokines can play a major role in progression of HCC and can increase liver injury in HCC patients (45). Among the cytokines, the most important one, IL-22 mainly secreted by TH17 is a member of IL10 cytokine family and plays pleiotropic protective and pathological roles in different tissues and organs. IL-22 affects hepatocytes and mucosal epithelial cells through specific receptors (46). Two main SNPs in the IL-22 gene including IL-22 rs1012356 A/T and rs1179251 C/G were found to be associated with HCV outcome (47).

5. *IL-18*

IL-18 induces interferon- γ and thus plays a critical role in activating the T-helper1 (TH1) which efficiently participates in antiviral immune response. The serum IL-18 level was found to be correlated with the severity of liver disease in cirrhotic patients, histological activity score and necrosis (48, 49).

Herein, some single nucleotide polymorphisms (SNPs) of IL18 gene were reported to be associated with spontaneous clearance of HCV infection (50).

6. *IL-28B*

A large literature has investigated spontaneous viral clearance and response to treatment associated with IL28B SNPs, located on chromosome 19 which encodes interferon lamda-3 (51-53). The SNPs of Rs809917, rs12979860 and rs12980275 are the main polymorphisms of IL28B gene, which are correlated with the prediction of response to therapy and spontaneous clearance (34, 54). These SNPs are often located in regulatory, transcription initiation or promoter region, which could impress on expression level of L28B cytokine and thereby decrease antiviral defense and spontaneous clearance of HCV infections. In genetic evolution, IL-28B gene includes different genotypes (SNPs) which are in favor of SVR in the HCV patients. Rs12979860 C/C genotype is associated with SVR in Caucasian, Asian, European, Hispanic and African-American populations. The genotype of rs8099917 has been reported to be the strongest predictor marker in genotype 2 and 3 (34, 48). Although rs8099917 GG/GG genotype indicated no SRV rate, the other polymorphism of the rs8099917 SNP like TG/GG allele had high SVR rate and spontaneous clearance in HCV patients in Asian population (5, 55-58). Another favorable IL28B polymorphism, commonly found in Asian population is CC genotype of rs12079860. The patients with CC polymorphism of rs12079860 SNP show spontaneous clearance and SVR rate 2-fold increase in comparison with the patients with TT genotype (56, 59). Additionally, those patients carrying rs12979860 SNP were associated with interferon-free treatment (33, 60, 61).

7. *Toll-like receptors (TLRs) genes*

Toll-like receptors (TLRs) are a type of proteins involved in innate immune system. TLRs are derived from the conserved structures with microbial origin. Some of the polymorphisms in TLR genes are found to be associated with viral infection. The TLR2, 4, 7, 8 and TLR9 are the most important elements related to antiviral responses. Several polymorphisms related to the genes of TLR4; rs498690A/G, rs4986791C/T, TLR7; rs179009A/G and TLR8; rs3764879 C/G play critical role on the susceptibility to HCV infection. Additionally, these TLRs can exert their impact on fibrosis progression and graft survival (55, 61-63).

8. *Suppressor of cytokine signaling 3 (SOCS3)*

SOCS3 is an inhibitor of IFN- α induced Janus kinase signal transducer and an activator of transcription pathways. SOCS3 polymorphisms influence on the

outcome of antiviral treatment. One polymorphism in -4874 site of SOCS3 gene with the ID name of rs4969170 A/G is the most important SNPs related to HCV therapy. Baseline of SOCS3 expression represents a novel blood biomarker which has the potential to predict the treatment response (26, 64).

9. PKR protein kinase

PKR or double-stranded RNA-dependent protein kinase is an interferon stimulating protein in the antiviral state. Some polymorphisms in PKR gene might affect on the treatment of HCV patients, who are under the interferon (INF) therapy. It was found that a polymorphism at position -168C/T in the promoter region of the PKR gene might be associated with natural outcome and also with response to IFN therapy in HCV infection. Additionally, tandem trinucleotide repeats in the promoter region of PKR can influence on the outcome of therapy. Consequently, up to nine of these repeats in PKR promoter were assigned to higher SVR (26, 65).

10. IL-1 β

IL-1 β is an initiating factor of inflammation cascade, which inhibits IFN- α/β and antiviral activity. When IL-1 β increases in the serum of HCV patients, consequently, the response to IFN-based therapy significantly decreases (66). Several SNPs have been reported to be in IL-1 β located at positions -31 (rs1143627), -511 (rs16944) and +3953 (rs1143634) which have high importance in HCV infection progression. Patients with genotype 4 with CC genotype have higher rate of SVR than those carrying CT and TT genotypes (26).

11. Myxovirus resistance A (MxA) gene

MxA gene is known as interferon stimulating gene, which encodes induced cytoplasmic interferon located in the human cells (67, 68). MxA protein inhibits the replication of many viruses which have negative-sense single-stranded RNA or which are double-stranded RNA viruses. Additionally, MxA might inhibit hepatitis B virus through suppressing viral reverse transcribing DNA (69). SNPs of MxA gene as important genetic host factors can influence on the outcome response to IFN- α in chronic HCV patients (70). Besides, antiviral drugs like ribavirin increases MxA gene expression in the patients (71). Additionally, some of intracellular antiviral proteins such as 2-5 oligoadenylate synthetase, dsRNA-activated protein kinase and MxA are induced by IFN and therefore have a defensive role against HCV infection. MxA protein expression as a sensitive marker for HCV replication could be a favorable predictor factor of SVR in the HCV patients with G4 treated with IFN- α 2 and ribavirin (13). An important

SNP, Rs2071430 located at position -88 MxA gene's promoter region with GG genotype is responsible for lowering MxA expression, whereas TT genotype of the same SNP is associated with protection against HCV infection (72). On the other hand, SNPs which are located on -88 (G/T) and -123 (C/A) of MxA gene promoter are associated with susceptibility to HCV infection and response to IFN- α as well as the natural outcome of hepatitis (68). Based on much literature, ribavirin induces an alternative MxA product with different functions, which acts as an immune-modulator in the pathway related to IFN- α antiviral activity (65, 71).

12. 2'-5'-oligoadenylate synthetase (2'-5'-OAS)

Oligoadenylate synthetase (2'-5'-OAS) stimulates IFN gene which plays a key role in viral RNA degradation and also inhibits viral replication. A polymorphism of 2'-5'-OAS gene with GG genotypes in 3' UTR region are known as non-responders to viral infection (73). However, several polymorphisms in 2'-5'-OAS gene are associated with susceptibility to HCV infection. A polymorphism of OAS1 gene, including rs2660 SNP located in the 3-untranslated region is found to be associated with the outcome of HCV infection (64).

13. IL-10

There are several polymorphisms in the IL-10 promoter, the most important of which include three SNPs; -1082G, -819 and -592. These SNPs have a major role on expression level of IL-10 gene. Therefore, lowering IL-10 expression drives immune system to Th1 type, which results in high-sustained viral clearance (66, 71). Most patients with low expression of IL10 are at high risk of rejection in liver transplantation whereas a polymorphism named RA-rs9610 A/G which lowers IL-10 expression shows a significant relation to HCV outcome (74).

14. TGF- β 1

Plasma level of TGF- β 1 is associated with liver fibrosis progression in HCV patients. TGF- β 1 polymorphisms at codon 10 (869) T/C and codon 25 (915) C/G of exon 1 can affect the chronicity of HCV disease and fibrotic progression (75-77). On the other hand, TGF- β 1 codon 25 SNP is strongly associated with SVR (76). Additionally, TGF- β 1 producing T cells have the ability to predict the absence of spontaneous clearance in patients with acute HCV infection (78).

15. Particular HLA

HLA alleles as host genetic factors show an efficient impact on susceptibility as well as the outcome of HCV treatment. For instance, frequencies of

DRB1*0401, DQB1*0301 and DQA1*0501 alleles in the patients infected chronically by HCV showed a significant association with the normal alanine aminotransferase (ALT) group leading the development of the chronic hepatitis C to liver damage and cirrhosis (79).

On the other hand, some of HLA alleles such as HLAB44, HLADR7, DRB1*0404, DRB1*0701, DRB1*11 and DQA1*0201-DQB1*02, DQA1*0501, DQB1*0301, DRB1*0401 alleles, DQA1*0301-DQB1*0301, DQA1*0501-DQB1*0301 and DRB1*1101-DQB1*0301 are significantly associated with raising SVR and lowering HCV infection; whereas DQB1*0201 allele is completely correlated with liver damage promotion in chronic HCV infection (77, 79).

16. Vitamin D receptors

One of the systemic hormones acts as multifunctional factors involved in immune response to microbial infections is vitamin D. This vitamin as an active form, including 1, 25-dihydroxy vitamin D, plays a critical role in immune regulation, cellular growth, programmed cell death and differentiation (80). The level of vitamin D in the body is considered as a predictor of response outcome related to treatment efficiency. High serum concentration of vitamin D is associated with non-SVR in HCV patients NR111 CCA haplotype. In addition, genetic variations in its receptor is associated with susceptibility to viral infection (79). Rs7975232CC SNP located on ApaI gene from VDR gene family contributes to non-SVR. Rs10877012 CC polymorphism in CYP27B1-1260 promoter is responsible for down-regulation of α -1-hydroxilation process and 25-OH vitamin D formation. This SNP reduces 1, 25 hydroxy vitamin D level and thus the reduced level of 1, 25 hydroxy vitamin D is associated with the failure of SVR in HCV patients with genotypes 1, 2 and 3. Besides, high level of vitamin D causes higher SVR rate which is associated with AA>CA polymorphism in CYP27B1-1260 pomoter gene (12).

17. Inosine Triphosphatase (ITPA)

Studies based on genome-wide analysis showed a significant association between ribavirin, hemolytic anemia and ITPA gene polymorphisms (57). Two important SNPs including Rs1127354 and rs7270101 in ITPA gene are considered as predictors of RBV-induced anemia. Several studies have reported that ITPA deficiency could reduce the rate of ribavirin-induced hemolytic anemia. For the HCV treatment, if taribavirin, which is a ribavirin analog with similar SVR, substituted ribavirin the rate of anemia would significantly decrease (9, 55, 57).

18. Intracellular adhesion molecules (ICAM)

ICAMs are known as cell surface proteins, which are expressed on hepatocytes. These membrane proteins are responsible for inflammation and hepatic damage in chronic HCV. Additionally, sICAM is secreted ICAM that is synthesized by various cell types. When more sICAM is released, there is increased the level of serum alanine aminotransferase that can progress the disease toward HCC in HCV patients. Increasing ICAM-1 expression promotes HCV infection and liver inflammation. Another polymorphism with identifier of Rs281437 located in ICAM-1 gene is strongly associated with progression of hepatic fibrosis in HCV genotype 4 (81).

19. Tumor necrosis factor- α (TNF- α)

TNF- α is type I cytokine which is secreted by monocytes and Kupffer cells. TNF- α is reported to be associated with fibrogenesis via binding to TNF-receptors. Polymorphisms in the TNF- α promoter seem to be related to histological severity at chronic HCV patients. TNF2 (-238A) and TNF3 (-308A) have polymorphisms in their promoter regions which are the cause of increased liver cirrhosis (82). Two other SNPs located in -1031C and -863A of promoter region of TNF- α are significantly associated with increasing the frequency of failure acute hepatitis (83).

20. DC-SIGNR binds to hepatitis C virus

The surface interaction between HCV and DC-SIGNR plays an important role in the pathogenesis of HCV infection. Tandem repeat polymorphisms located within DC-SIGNR gene have been recently reported to be associated with HCV replication. Decreased attachment of DC-SIGNR to HCV might be associated with lower HCV replication. Therefore, the patients carrying less number of tandem repeats, approximately lower than four repeats, have been observed to be less compatible host for HCV replication than those carrying up to nine repeats (84).

21. Apolipoprotein E (APOE)

APOE as a component of VLDL binds to receptors, which HCV used for entering into the cells. There are three isoforms of APOE including: APOE2, APOE3 and APOE4. APOE2 allele typically binds to lipoviral particle and acts as a barrier to the entry of the HCV to cells (85, 86).

22. Osteopontin (OPN) Gene

Osteopontin Gene expresses a secretory protein, which strongly attaches to osteoclasts through hydroxylapatite in the bone matrix. The presence of polymorphisms in OPN gene affect IFN-based therapies in HCV chronic patients. The SNP at the

nucleotide -443 C/T versus C/C and T/T is a favorable predictor for the response to interferon therapy in chronic HCV patients (87).

Discussion

Interferon type I was used as a first line of treatment for HCV infection in 1986. Afterward, ribavirin was added to therapy regime of HCV patients. Aimed at providing more efficient treatment, pegylated interferon -alfa (PEG-IFN- α) entered to the practical cure of HCV patients in 2001 and its combination with ribavirin was considered as the standard common medicine (69, 88, 89). The therapy regime with the combination of PEG-IFN and ribavirin showed a significant increase in SVR in comparison with conventional therapy in which IFN- α and/or ribavirin were administered separately (5). Telaprevir and Boceprevir were introduced in 2011, which were prescribed as a triple therapy for HCV patients (90). Two other drugs, Sofosbuvir and Simeprevir, which act as inhibitor of NS5B nucleotide polymerase and NS3/4 protease have been also used for HCV treatment since 2013 (34). Faldaprevir and Daclatasvir as new direct-acting antiviral (DAA) medicine were expected to be available in 2015 (91). In spite of the recent therapeutic success with standard-of-care treatment (SOC), several patients are annually reported that unequally respond to the therapy regime. Based on clinical reports, it is evidenced that a combination regime including Peg-IFN and ribavirin plus telaprevir can increase SVR rate of hepatitis C patients with genotype 1 (92). It is expected that new efficient strategies for the treatment of hepatitis C virus infection, may lead to simplified medications with new pharmaceutical drugs. Up to now, scientists are hopeful to develop more specific drug regimes for inhibiting HCV infection and to optimize drugs administration based on individual genetic variations in order to decrease medication side effects in the near future (5).

According to the growing trend of new hepatitis medications, administration of interferon-free regime can be predicted. A combination of faldaprevir and deleobuvir with ribavirin as an interferon-free regime results in 59% SVR rate in HCV patients and also reduces the anemia due to the absence of PEG-IFN adverse effects on the white cells and platelets (92).

The treatment of HCV is expensive and associated with adverse consequences for patients and providers will need to know the likelihood of the response to treatment. The factors which determine this likelihood are called pretreatment response and can be classified into host, viral and environmental factors. Multiple genes are affected when an individual is infected by HCV due to various interactions which occurs between viral and host

factor genes. Therefore, the response to HCV infection in different HCV positive individuals strongly depends on varying factors such as age, race, insulin resistance, obesity, co-infection with EBV, HIV and HBV, drugs, steatosis, alcohol consumption, hyperlipidemia, diabetes and iron (7, 34, 85, 93-95). Despite many advances for treating and recovering the HCV infected patients, most health professionals are poorly aware of the new strategies to enhance HCV assessment and treatment [100]. Although, the prediction outcome of response is not exactly satisfying, most laboratories trails need to achieve the appropriate predictors. For instance, occasionally, biomarkers used for detection of early HCC can alter treatment decision and improve patient's survival. In addition, several gene polymorphisms, which are related to immune response and influence the outcome of acute and chronic HCV infection, facilitate the response prediction.

Conclusion

Recently, combination of PEG-IFN- α and ribavirin is considered as the standard and common treatment for HCV. Although some medications have emerged for improving the treatment regime of HCV, none of them shows the same efficiency in all patients. Therefore, it seems that there is no practical medication for preventing HCV infection worldwide. Taken together, HCV treatment has many limitations, which host genetic factors are most important causes of progression of HCV infection. The most efficient strategy for HCV medications is to drive immune response towards cellular immune system which activates TH1 and directs the disease to acute infection. Considering that acute infectious hepatitis shows high SVR and viral clearance, a favorable medication can improve HCV treatment for all patients. Analysis of gene polymorphisms can help to better understanding of interactions of host and virus and may consequently lead to prediction of the sustained virological response to antiviral therapy. Besides, those patients with adverse alleles may benefit from additional treatment strategies.

Author Contributions

All authors have the same contribution in preparation of data and manuscript writing.

Conflict of Interest

There is no conflict of interest in this review article.

References

1. Ashtari S, Vahedi M, Pourhoseingholi MA, Karkhane M, Kimia Z, Pourhoseingholi A, et al. Direct medical care costs associated with patients diagnosed with chronic HCV. *Hepat Mon.* 2013; 13(5): e8415. PMID: 23930132

2. Foster GR, Coppola C, Derbala M, Ferenci P, Orlandini A, Reddy KR, et al. Impact of safety-related dose reductions or discontinuations on sustained virologic response in HCV-infected patients: Results from the GUARD-C Cohort. *PloS one*. 2016; 11(3): e0151703. PMID: 4809570
3. Zhang T, Li Y, Ho WZ. Drug abuse, innate immunity and hepatitis C virus. *Rev Med virol*. 2006; 16(5): 311-27. PMID: 16933366
4. Gorodin S, Unal S, Wang Y, Mikhaylov MI, Bigbulatova L, Jehuda-Cohen T. New tools in HCV diagnosis, in light of the enhanced awareness and the new drugs for treatment: SMARTube and stimmunology. *Scientific World J*. 2013(2013): ID389780. PMID: 23476130
5. Chuang WL, Yu ML. Host factors determining the efficacy of hepatitis C treatment. *J Gastroenterol*. 2013;48(1):22-30. PMID: 23104468
6. Alavian SM, Adibi P, Zali MR. Hepatitis C virus in Iran: Epidemiology of an emerging infection. *Arch Iranian Med*. 2005; 8(2):84-90.
7. Rezaee-Zavareh MS, Hesamizadeh K, Sharafi H, Alavian SM. Treatment of chronic hepatitis C infection with Direct-acting Antiviral Agents in Liver-Transplant patients: A Systematic Review and Meta-Analysis. *Hepat mon*. 2017; 17(6):e12324.
8. Ramos JA, Silva R, Hoffmann L, Ramos ALA, Cabello PH, Úrményi TP, et al. Association of IL-10, IL-4, and IL-28B gene polymorphisms with spontaneous clearance of hepatitis C virus in a population from Rio de Janeiro. *BMC Res Notes*. 2012; 5:508. PMID: 22976179
9. Rau M, Baur K, Geier A. Host genetic variants in the pathogenesis of hepatitis C. *Viruses*. 2012; 4(12):3281-302. PMID: 23342360
10. Singh S, Singh PP, Roberts LR, Sanchez W. Chemopreventive strategies in hepatocellular carcinoma. *Nat Rev Gastroenterol Hepatol*. 2014; 11(1):45-54. PMID: 23938459
11. Wantuck J, Ahmed A, Nguyen M. Review article: the epidemiology and therapy of chronic hepatitis C genotypes 4, 5 and 6. *Aliment Pharmacol Ther*. 2014; 39(2):137-47. PMID: 24251930
12. Suruki RY, Mueller N, Hayashi K, Harn D, DeGruttola V, Raker CA. Host immune status and incidence of hepatocellular carcinoma among subjects infected with hepatitis C virus: a nested case-control study in Japan. *Cancer Epidemiol Biomarkers Prev*. 2006; 15(12):2521-5. PMID: 17164379
13. Shaker O, Ahmed A, Doss W, Abdel-Hamid M. Mx_A Expression as marker for assessing the therapeutic response in HCV genotype 4 Egyptian patients. *J Viral Hepat*. 2010; 17(11):794-9. PMID: 20002306
14. Van-Lume DSdM, de Albuquerque MdFP, de Souza AI, Domingues ALC, Lopes EPda, de Moraes CNL. Association between Schistosomiasis mansoni and hepatitis C: systematic review. *Rev Saúde Pública*. 2013; 47(2):414-24. PMID: 24057369
15. Reed K, Rice C. Overview of hepatitis C virus genome structure, polyprotein processing, and protein properties. *The Hepatitis C Viruses*: NewYork: Springer; 2000. p. 55-84.
16. Younossi ZM, Baranova A, Afendy A, Collantes R, Stepanova M, Manyam G. Early gene expression profiles of patients with chronic hepatitis C treated with pegylated interferon-alfa and ribavirin. *Hepatol*. 2009;49(3):763-74. PMID: 19140155
17. Vivithanaporn P, Maingat F, Lin LT, Na H, Richardson CD, Agrawal B. Hepatitis C virus core protein induces neuroimmune activation and potentiates Human Immunodeficiency Virus-1 neurotoxicity. *PLoS One*. 2010; 5(9):e12856. PMID: 20877724
18. Shiryayev SA, Thomsen ER, Cieplak P, Chudin E, Cheltsov AV, Chee MS. New details of HCV NS3/4A proteinase functionality revealed by a high-throughput cleavage assay. *PLoS One*. 2012; 7(4):e35759. PMID: 22558217
19. Dumont S, Cheng W, Serebrov V, Beran RK, Tinoco I, Pyle AM. RNA translocation and unwinding mechanism of HCV NS3 helicase and its coordination by ATP. *Nature*. 2006; 439(7072):105-8. PMID: 16397502
20. Woodhouse SD, Narayan R, Latham S, Lee S, Antrobus R, Gangadharan B, et al. Transcriptome sequencing, microarray, and proteomic analyses reveal cellular and metabolic impact of hepatitis C virus infection in vitro. *Hepatol*. 2010;52(2):443-53. PMID: 20683944
21. Abdel-Hakeem MS, Shoukry NH. Protective immunity against hepatitis C: many shades of gray. *Frontiers in immunology*. 2014; 5:274. PMID: 24982656
22. Schinazi R, Halfon P, Marcellin P, Asselah T. HCV direct-acting antiviral agents: the best interferon-free combinations. *Liver Int*. 2014; 34(s1):69-78. PMID: 24373081
23. Ashtari S, Vahedi M, Pourhoseingholi MA, Pourhoseingholi A, Safaee A, Moghimi-Dehkordi B, et al. Estimation of average diagnosis and treatment costs of hepatitis C. *Gastroenterol Hepatol Bed Bench*. 2012; 5(3):139-45. PMID: 24834215
24. Gerlach JT, Diepolder HM, Zachoval R, Gruener NH, Jung M-C, Ulsenheimer A, et al. Acute hepatitis C: high rate of both spontaneous and treatment-induced viral clearance. *Gastroenterol*. 2003; 125(1):80-8. PMID: 12851873
25. Ye L, Peng JS, Wang X, Wang YJ, Luo GX, Ho WZ. Methamphetamine enhances Hepatitis C virus replication in human hepatocytes. *J viral hepat*. 2008; 15(4):261-70. PMID: 18307590.
26. Omran MH, Ibrahim NE, Youssef SS, Fatouh BE, Nabil W, El-Shami MM, et al. Relation of interleukin-1 β gene to treatment response in chronic patients infected with HCV genotype 4. *The J Infect Dev Ctries*. 2013; 7(11):851-8. PMID: 24240044
27. Moghadam FS, Mohebbi SR, Hosseini SM, Damav B, Zali MR. A new subtype of hepatitis C virus genotype 3: Analysis of available evidence. *Hepat Mon*. 2013; 13(8). E13380. PMID: 17935166
28. Messina JP, Humphreys I, Flaxman A, Brown A, Cooke GS, Pybus OG, et al. Global distribution and prevalence of hepatitis C virus genotypes. *Hepatol*. 2015; 61(1):77-87. PMID: 25069599
29. Shiffman ML, Di Bisceglie AM, Lindsay KL, Morishima C, Wright EC, Everson GT, et al. Peginterferon Alfa-2a and ribavirin in patients with chronic hepatitis C who have failed prior treatment. *Gastroenterol*. 2004; 126(4):1015-23. PMID: 15059949
30. Sarrazin C, Susser S, Doehring A, Lange CM, Müller T, Schleckner C, et al. Importance of IL28B gene polymorphisms in hepatitis C virus genotype 2 and 3 infected patients. *J Hepatol*. 2011; 54(3):415-21. PMID: 21112657

31. Zeuzem S, Soriano V, Asselah T, Bronowicki J-P, Lohse AW, Müllhaupt B, et al. Faldaprevir and ledipasvir for HCV genotype 1 infection. *N Engl J Med*. 2013; 369(7):630-9. PMID: 23944300
32. Jensen DM, Brunda M, Elston R, Gane EJ, George J, et al. Interferon-free Regimens containing Setrobuvir for Patients with Genotype 1 Chronic Hepatitis C: A randomized, Multicenter study. *Liver Int*. 2016;36(4): 505-14. PMID: 26519669
33. Sulkowski M, Rodriguez-Torres M, Lawitz E, Shiffman M, Pol S, Herring R, et al. 1421 High Sustained Virologic Response Rate in Treatment-Naive HCV Genotype 1a and 1b Patients Treated for 12 Weeks with an Interferon-Free All-Oral Quad Regimen: Interim Results. *J Hepatol*. 2012; 56:S560.
34. Jimenez-Sousa MA, Fernandez-Rodriguez A, Guzman-Fulgencio M, Garcia-Alvarez M, Resino S. Meta-analysis: implications of interleukin-28B polymorphisms in spontaneous and treatment-related clearance for patients with hepatitis C. *BMC Med*. 2013;11:6. PMID: 23298311
35. Moura AS, Carmo RA, Teixeira AL, Leite VH, Rocha MO. Soluble inflammatory markers as predictors of liver histological changes in patients with chronic hepatitis C virus infection. *European journal of clinical microbiology & infectious diseases: official publication of the European Society of Clinical Microbiology*. 2010; 29(9):1153-61. PMID: 20559676
36. Karkhane M, Mohebbi SR, Azimzadeh P, Niasar MS, Sarbazi MR, Sharifian A, et al. Lack of association between interleukin 28B gene polymorphisms (rs8099917G/T, rs12979860 C/T) and susceptibility to chronic hepatitis C virus infection, Tehran, Iran. *Gastroenterol Hepatol Bed Bench*. 2016; 9(S11):S29. PMID: 28224025
37. Azimzadeh P, Mohebbi SR, Romani S, Kazemian S, Mirtalebi H, Vahedi M, et al. Role of TGF- β 1 codon 10 polymorphism in chronic hepatitis C patients. *J Babol Univ Med Sci*. 2011; 13(4).
38. Romani S, Azimzadeh P, Mohebbi SR, Kazemian S, Almasi S, Naghoosi H, et al. Investigation of transforming growth factor- β 1 gene polymorphisms among Iranian patients with chronic hepatitis C. *Hepat Mon*. 2011;11(11):901-6. PMID: 22363353
39. Salehi Moghadam F, Mohebbi SR, Hosseini SM, Romani S, Mirtalebi H, Azimzadeh P, et al. Phylogenetic analysis of hepatitis C virus strains and risk factors associated with infection and viral subtypes among Iranian patients. *J Med Virol*. 2014; 86(8):1342-9. PMID: 24838700
40. Ashtari S, Vahedi M, Pourhoseingholi MA, Zali MR. Evaluated outcomes in patients with chronic hepatitis C. *Gastroenterol Hepatol Bed Bench*. 2013; (S6):S58-S64.
41. Fan X, Liu W, Li C, Wang Z, Luo L, Tan D, et al. Circulating Th1 and Th2 cytokines in patients with hepatitis C virus infection. *Mediators of inflammation*. 1998; 7(4):295-7. PMID: 9792342
42. Polyak SJ, Khabar KS, Rezeiq M, Gretch DR. Elevated levels of interleukin-8 in serum are associated with hepatitis C virus infection and resistance to interferon therapy. *J Virol*. 2001; 75(13):6209-11. PMID: 11390624
43. Masumoto T, Ohkubo K, Yamamoto K, Ninomiya T, Abe M, Akbar S, et al. Serum IL-8 levels and localization of IL-8 in liver from patients with chronic viral hepatitis. *Hepato-gastroenterology*. 1997; 45(23):1630-4. PMID: 9840119
44. Jahan S, Khaliq S, Ijaz B, Ahmad W, Hassan S. Role of HCV Core gene of genotype 1a and 3a and host gene Cox-2 in HCV-induced pathogenesis. *Virology*. 2011; 8:155. PMID: 21457561
45. Sousa GM, Oliveira IS, Andrade LJ, Sousa-Atta ML, Parana R, Atta AM. Serum levels of Th17 associated cytokines in chronic hepatitis C virus infection. *Cytokine*. 2012; 60(1):138-42. PMID: 22748467
46. Hennig BJ, Frodsham AJ, Hellier S, Knapp S, Yee LJ, Wright M, et al. Influence of IL-10RA and IL-22 polymorphisms on outcome of hepatitis C virus infection. *Liver Int*. 2007; 27(8):1134-43. PMID: 17845543
47. Huang CF, Yeh ML, Hsieh MH, Hsieh MY, Lin ZY, Chen SC, et al. Clinical utility of host genetic IL-28B variants in hepatitis C virus genotype 1 Asian patients retreated with pegylated interferon plus ribavirin. *J Gastroenterol Hepatol*. 2013; 28(9):1515-20. PMID: 23560893
48. Chen TY, Hsieh YS, Wu TT, Yang SF, Wu CJ, Tsay GJ, et al. Impact of serum levels and gene polymorphism of cytokines on chronic hepatitis C infection. *Translational research: J lab clin Med*. 2007; 150(2):116-21. PMID: 17656331
49. Sharma A, Chakraborti A, Das A, Dhiman RK, Chawla Y. Elevation of interleukin-18 in chronic hepatitis C: implications for hepatitis C virus pathogenesis. *Immunology*. 2009; 128(1pt2):e514-e22. PMID: 19740312
50. Chattergoon MA, Levine JS, Latanich R, Osburn WO, Thomas DL, Cox AL. High plasma interleukin-18 levels mark the acute phase of hepatitis C virus infection. *J Infect Dis*. 2011; 204(11):1730-40. PMID: 21984735
51. Mizokami M. Discovery of critical host factor, IL-28B, associated with response to hepatitis C virus treatment. *J Gastroenterol Hepatol*. 2012; 27(3):425-9. PMID: 22168813
52. Arends JE, Franssen JH, Hoepelman AI, van Baarle D. Association between IL28B polymorphisms and first-phase viral load decrease in chronic hepatitis C virus-infected patients treated with peginterferon alfa-2b/ribavirin. *Int J Antimicrob Agents*. 2011; 38(6):538-9. PMID: 21974859
53. Ciesla A, Bociaga-Jasik M, Sobczyk-Krupiarz I, Glowacki MK, Owczarek D, Cibor D, et al. IL28B polymorphism as a predictor of antiviral response in chronic hepatitis C. *World J Gastroenterol*. 2012; 18(35):4892-7. PMID: 23002361
54. Fonseca-Coronado S, Vaughan G, Cruz-Rivera MY, Carpio-Pedroza JC, Ruiz-Tovar K, Ruiz-Pacheco JA, et al. Interleukin-28B genotyping by melt-mismatch amplification mutation assay PCR analysis using single nucleotide polymorphisms rs12979860 and rs8099917, a useful tool for prediction of therapy response in hepatitis C patients. *J Clin Microbiol*. 2011; 49(7):2706-10. PMID: 21613433
55. Cariani E, Villa E, Rota C, Critelli R, Trenti T. Translating pharmacogenetics into clinical practice: interleukin (IL) 28B and inosine triphosphatase (ITPA) polymorphisms in hepatitis C virus (HCV) infection. *Clin Chem Lab Med*. 2011; 49(8):1247-56. PMID: 21612542
56. Pagliaccetti NE, Robek MD. Interferon-lambda in HCV Infection and Therapy. *Viruses*. 2010; 2(8):1589-602. PMID: 21994696
57. Di Marco V, Calvaruso V, Grimaudo S, Ferraro D, Pipitone R, Di Stefano R, et al. Role of IL-28B and inosine triphosphatase polymorphisms in efficacy and safety of Peg-Interferon and ribavirin in chronic hepatitis C compensated cirrhosis with and

- without oesophageal varices. *J Viral Hepat.* 2013; 20(2):113-21. PMID: 23301546
58. Poordad F, Lawitz E, Kowdley K, Everson G, Freilich B, Cohen D, et al. 1399 12-week interferon-free regimen of abt-450/r+ abt-333+ ribavirin achieved svr12 in more than 90% of treatment-naïve hcv genotype-1-infected subjects and 47% of previous non-responders. *J Hepatol.* 2012; 56(S5):49-S50.
59. Patel K, Lucas JE, Thompson JW, Dubois LG, Tillmann HL, Thompson AJ, et al. High predictive accuracy of an unbiased proteomic profile for sustained virologic response in chronic hepatitis C patients. *Hepatol.* 2011; 53(6):1809-18. PMID: 21381069
60. Neumann U. Impact of IL-28B polymorphism on outcome in patients with hepatitis C after liver transplantation. *Expert Rev Gastroenterol Hepatol.* 2011; 5(4):429-31. PMID: 21780887
61. Berger CT, Kim AY. IL28B polymorphisms as a pretreatment predictor of response to HCV treatment. *Infect Dis Clinics North America.* 2012; 26(4):863-77. PMID: 23083820
62. Al-Qahtani AA, Al-Anazi MR, Al-Zoghaibi F, Abdo AA, Sanai FM, Khan MQ, et al. The association of toll-like receptor 4 polymorphism with hepatitis C virus infection in Saudi Arabian patients. *BioMed Res Int.* 2014; 2014: 357062. PMID: 25177689
63. Persico M, Capasso M, Russo R, Persico E, Croce L, Tiribelli C, et al. Elevated expression and polymorphisms of SOCS3 influence patient response to antiviral therapy in chronic hepatitis C. *Gut.* 2008; 57(4):507-15. PMID: 17881539
64. Knapp S, Yee L, Frodsham A, Hennig B, Hellier S, Zhang L, et al. Polymorphisms in interferon-induced genes and the outcome of hepatitis C virus infection: roles of MxA, OAS-1 and PKR. *Genes Immun.* 2003; 4(6):411-9. PMID: 12944978
65. Gao B, Hong F, Radaeva S. Host factors and failure of interferon- α treatment in hepatitis C virus. *Hepatol.* 2004; 39(4):880-90. PMID: 15057887
66. Glymph S, Mandal S, Knowell AE, Abebe F, Chaudhary J. The myxovirus resistance A (MxA) gene- 88G> T single nucleotide polymorphism is associated with prostate cancer. *Infect, Genet Evol.* 2013; 16:186-90. PMID: 23438650
67. Hijikata M, Mishiro S, Miyamoto C, Furuichi Y, Hashimoto M, Ohta Y. Genetic polymorphism of the MxA gene promoter and interferon responsiveness of hepatitis C patients: revisited by analyzing two SNP sites (-123 and-88) in vivo and in vitro. *Intervirology.* 2002; 44(6):379-82. PMID: 11805446
68. Netherton CL, Simpson J, Haller O, Wileman TE, Takamatsu H-H, Monaghan P, et al. Inhibition of a large double-stranded DNA virus by MxA protein. *J Virol.* 2009; 83(5):2310-20. PMID: 19109387
69. Sadeq R, Mohtady H, Al Badawy N, Ibrahim S, Omar AR, Husseiny MI, et al. Endogenous IFN γ in chronic HCV genotype 4 patients treated with PEG-IFN α and ribavirin. *J Infect Dev Ctries.* 2013; 7(11):859-67. PMID: 24240045
70. Stevenson NJ, Murphy AG, Bourke NM, Keogh CA, Hegarty JE, O'Farrelly C. Ribavirin enhances IFN- α signalling and MxA expression: a novel immune modulation mechanism during treatment of HCV. *PLoS One.* 2011; 6(11):e27866. PMID: 22114715
71. Abbas Z, Moatter T, Hussainy A, Jafri W. Effect of cytokine gene polymorphism on histological activity index, viral load and response to treatment in patients with chronic hepatitis C genotype 3. *World J Gastroenterol.* 2005; 11(42):6656. PMID: 16425360
72. Suppiah V, Moldovan M, Ahlenstiel G, Berg T, Weltman M, Abate ML, et al. IL28B is associated with response to chronic hepatitis C interferon- α and ribavirin therapy. *Nat Genet.* 2009; 41(10):1100-4. PMID: 19749758
73. Taguchi T, Nagano-Fujii M, Akutsu M, Kadoya H, Ohgimoto S, Ishido S, et al. Hepatitis C virus NS5A protein interacts with 2', 5'-oligoadenylate synthetase and inhibits antiviral activity of IFN in an IFN sensitivity-determining region-independent manner. *J Gen Virol.* 2004; 85(4):959-69. PMID: 15039538
74. Suzuki S, Tanaka Y, Orito E, Sugauchi F, Hasegawa I, Sakurai M, et al. Transforming growth factor-beta-1 genetic polymorphism in Japanese patients with chronic hepatitis C virus infection. *J Gastroenterol Hepatol.* 2003; 18(10):1139-43. PMID: 12974889
75. Pereira FA, Pinheiro da Silva NN, Rodart IF, Carmo TM, Lemaire DC, Reis MG. Association of TGF-beta1 codon 25 (G915C) polymorphism with hepatitis C virus infection. *J Med virol.* 2008; 80(1):58-64. PMID: 18041006
76. Gewaltig J, Mangasser-Stephan K, Gartung C, Biesterfeld S, Gressner AM. Association of polymorphisms of the transforming growth factor- β 1 gene with the rate of progression of HCV-induced liver fibrosis. *Clin Chim Acta.* 2002; 316(1):83-94. PMID: 11750277
77. Harfouch S, Guiguet M, Valantin M-A, Samri A, Ouazene Z, Slama L, et al. Lack of TGF- β production by hepatitis C virus-specific T cells during HCV acute phase is associated with HCV clearance in HIV coinfection. *J Hepatol.* 2012; 56(6):1259-68. PMID: 22326469
78. Hill AV. The immunogenetics of human infectious diseases. *Ann Rev Immun.* 1998; 16(1):593-617. PMID: 9597143
79. Yu RB, Hong X, Ding WL, Tan YF, Zhang YX, Sun NX, et al. The association between the genetic polymorphism of HLA-DQA1, DQB1, and DRB1 and serum alanine aminotransferase levels in chronic hepatitis C in the Chinese population. *J Gastroenterol Hepatol.* 2008; 23(9):1394-402. PMID: 18028350
80. Mahmoudi T, Mohebbi SR, Pourhoseingholi MA, Fatemi SR, Zali MR. Vitamin D receptor gene ApaI polymorphism is associated with susceptibility to colorectal cancer. *Dig Dis sci.* 2010; 55(7):2008-13. PMID: 19795209
81. Rizk NM, Derbala MF. Genetic polymorphisms of ICAM 1 and IL28 as predictors of liver fibrosis severity and viral clearance in hepatitis C genotype 4. *Clin Res Hepatol Gastroenterol.* 2013; 37(3):262-8. PMID: 23137758
82. Höhler T, Kruger A, Gerken G, Schneider PM, zum Büschenfelde KHM, Rittner C. Tumor necrosis factor alpha promoter polymorphism at position-238 is associated with chronic active hepatitis C infection. *J Med virol.* 1998; 54(3):173-7. PMID: 9515764
83. Nattermann J, Ahlenstiel G, Berg T, Feldmann G, Nischalke H, Müller T, et al. The tandem-repeat polymorphism of the DC-SIGNR gene in HCV infection. *J viral hepat.* 2006; 13(1):42-6. PMID: 163644081
84. Price DA, Bassendine MF, Norris S, Golding C, Toms GL, Schmid ML, et al. Apolipoprotein ϵ 3 allele is associated with

persistent hepatitis C virus infection. *Gut*. 2006; 55(5):715-8. PMID: 16299033

85. Minakari M, Sameni FK, Shalmani HM, Molaee M, Zali MR. Hepatic steatosis in Iranian patients with chronic hepatitis C. *Med Princ Pract*. 2008; 17(2):126-30. PMID: 18288796

86. Naito M, Matsui A, Inao M, Nagoshi S, Nagano M, Ito N, et al. SNPs in the promoter region of the osteopontin gene as a marker predicting the efficacy of interferon-based therapies in patients with chronic hepatitis C. *J Gastroenterol*. 2005; 40(4):381-8. PMID: 15868370

87. Shaker O, El-Shehaby A, Fayez S, Zahra A, Marzouk S, El Raziky M. Osteopontin gene polymorphisms as predictors for the efficacy of interferon therapy in chronic hepatitis C Egyptian patients with genotype 4. *Cell Biochem Funct*. 2013; 31(7):620-5. PMID: 23400862

88. Zali MR, Mayumi M, Raoufi M, Nowroozi A. Hepatitis C virus genotypes in the Islamic Republic of Iran: A preliminary study. *East Mediterr Health J*. 2000; 6(2-3):372-7. PMID: 11556026

89. Yamada G, Iino S, Okuno T, Omata M, Kiyosawa K, Kumada H, et al. Virological response in patients with hepatitis C virus genotype 1b and a high viral load: impact of peginterferon-alpha-2a plus ribavirin dose reductions and host-related factors. *Clin Drug Investig*. 2008; 28(1):9-16. PMID: 18081356

90. Goralczyk AD, Cameron S, Amanzada A. Treatment of chronic HCV genotype 1 infection with telaprevir: a Bayesian mixed treatment comparison of fixed-length and response-guided treatment regimens in treatment-naïve and-experienced patients. *BMC Gastroenterol*. 2013; 13(1):148. PMID: 24118976

91. Olmstead AD, Knecht W, Lazarov I, Dixit SB, Jean F. Human subtilase SKI-1/S1P is a master regulator of the HCV Lifecycle and a potential host cell target for developing indirect-acting antiviral agents. *PLoS Pathogens*. 2012; 8(1):e1002468. PMID: 22241994

92. Zeuzem S, Andreone P, Pol S, Lawitz E, Diago M, Roberts S, et al. Telaprevir for retreatment of HCV infection. *N Engl J Med*. 2011; 364(25):2417-28. PMID: 21696308

93. Moura AS, Carmo RA, Teixeira AL, Teixeira MM, Rocha MO. Soluble inflammatory markers as predictors of virological response in patients with chronic hepatitis C virus infection treated with interferon-alpha plus ribavirin. *Memorias do Instituto Oswaldo Cruz*. 2011; 106(1):38-43. PMID: 21340353

94. Tahaei SME, Mohebbi SR, Azimzadeh P, Vahedi M, Almasi S, Romani S, et al. Frequency of HIV and HCV co-infections in chronic HBV patients referred to Taleghani Hospital, Tehran, Iran from 2006 to 2010. *Hepat Mon*. 2011; 11(12):993-6.

95. Rezaei-Tavirani M, Safaei A, Zali MR. The association between polymorphisms in insulin and obesity related genes and risk of colorectal cancer. *Iran J Cancer Prev*. 2013; 6(4):179-85. PMID: 25250132